Determination of tonicity effects of ketoacids and lactate by use of two canine red blood cell assays

Jennifer M. Reinhart DVM, MS
Misty R. Yancey BS
Jennifer D. Girard-Denton DVM
Thomas Schermerhorn VMD

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From the Department of Clinical Sciences, College of Veterinary Medicine, Kansas State University, Manhattan, KS 66506. Dr. Reinhart’s present address is Department of Medical Sciences, School of Veterinary Medicine, University of Wisconsin, Madison, WI 53706. Dr. Girard-Denton’s present address is Banfield Pet Hospital, 5 Mystic View Rd, Everett, MA 02149.

Address correspondence to Dr. Reinhart (jreinhart@wisc.edu).

The terms osmolality and tonicity are often used interchangeably; however, these 2 concepts differ in subtle yet important ways. Osmolality describes the total solute concentration but does not distinguish between solutes that exert osmotic pressure (effective osmoles) and those that do not (ineffective osmoles). The barrier function of a cell membrane is an essential component in the generation of an osmotic gradient, and the ability to exert osmotic pressure depends on the permeability of a cell membrane to a given osmole. Tonicity, also called to define hypertonicity. In a cross-sectional study,6 the prevalence of hypertonicity in human diabetics differs depending on the cutoff used.

Major contributors to serum tonicity include the monovalent ions and glucose. In many disease states, hypertonicity results from an increased serum concentration of endogenous or exogenous effective osmoles. Diabetes mellitus is an important cause of hypertonicity, which results from hyperglycemia, in humans and domestic animals. The prevalence of hypertonicity in human diabetics differs depending on the cutoff used to define hypertonicity. In a cross-sectional study,7 the prevalence in human diabetics was reported as 35% to 78%; a similar range has been reported in diabetic dogs and cats (35% to 81%).

In addition to the glucose concentration, the sodium concentration also appears to be an important contributor to hypertonicity in human and veterinary patients and may independently impact diabetic progression.9 Diabetic patients may also have excess accumulation of endogenous osmoles that typically are maintained at minute concentrations in healthy patients. Examples of these solutes are the ketoacids β-hydroxybutyrate and acetoacetate, which are overproduced in unregulated pathological hypertonicity will disrupt these pathways.4 Finally, cells acclimatized to chronic hypertonicity can swell during rapid correction of serum tonicity, which leads to tissue edema.5

For both assays, β-hydroxybutyrate and acetoacetate acted as ineffective osmoles, whereas lactate acted as an effective osmole in 3 of 4 experiments. (Am J Vet Res 2015;76:77–83)

ABBREVIATION

H50  Sucrose concentration at which 50% hemolysis is detected

OBJECTIVE

To determine the tonicity effects of β-hydroxybutyrate, acetoacetate, and lactate in canine RBCs.

SAMPLE

RBCs from approximately 40 dogs.

PROCEDURES

2 in vitro methods were used to conduct 4 experiments. The modified osmotic fragility assay was used to measure the ability of ketoacid salts added to serial sucrose dilutions to protect RBCs from osmotic hemolysis. In a second assay, a handheld cell counting device was used to measure changes in RBC diameter to assess the tonicity effect of solutions of ketoacid and lactate salts.

RESULTS

For the modified osmotic fragility assay, all ketoacid salts had an osmoprotective effect, but the effect was determined to be completely attributable to the tonicity effect of added cations (sodium and lithium) and not the ketoacid moieties. However, both the sodium and lithium lactate salts provided osmoprotection attributable to both the cation and lactate anion. For the second assay, RBC diameter was significantly increased with the addition of urea (an ineffective osmole) but did not change with the addition of glucose (an effective osmole), which established the behaviors of ineffective and effective osmoles in this assay. The RBC diameter was significantly increased over that of control samples by the addition of sodium β-hydroxybutyrate, lithium acetoacetate, and lithium lactate but was decreased by the addition of sodium lactate.

CONCLUSIONS AND CLINICAL RELEVANCE

For both assays, β-hydroxybutyrate and acetoacetate acted as ineffective osmoles, whereas lactate acted as an effective osmole in 3 of 4 experiments. (Am J Vet Res 2015;76:77–83)
diabetes mellitus by the liver as an indirect consequence of relative or absolute insulin deficiency. Serum lactate concentration can also increase, particularly during diabetic ketoacidosis, owing to hypoperfusion and resultant anerobic glycolysis.

These solutes increase serum osmolality, but it is unclear whether they increase serum tonicity by acting as effective osmoles. Investigators in 1 study evaluated the effects of glucose, with or without ketoacids, on a number of canine RBC variables, including size and osmotic fragility. They found that changes in these variables are significantly different when both glucose and sodium β-hydroxybutyrate are added, compared with changes after the addition of glucose alone. By use of an osmotic fragility assay with human RBCs, investigators in another study concluded that acetoacetate is an effective osmole because lithium acetoacetate protects RBCs from hypotonic-induced lysis in a manner similar to that of the known effective osmoles NaCl and glucose. However, the methods used in previous investigations involved the addition of ketoacids to solutions in the form of sodium or lithium salts of the weak organic acid, and tonicity effects of added cations may have confounded the analysis. Thus, the tonicity effects of organic anions remain to be firmly established.

The purpose of the study reported here was to determine whether organic anions are effective osmoles. We hypothesized that β-hydroxybutyrate, acetoacetate, and lactate would be ineffective osmoles and would not exert a tonicity effect on canine RBCs.

Materials and Methods

SAMPLES

Surplus EDTA-anticoagulated blood from approximately 40 dogs, stored at 4°C, was obtained within 48 hours after collection; samples were obtained from the Kansas State University Veterinary Diagnostic Laboratory Clinical Pathology Laboratory after routine hematologic analysis. No samples were collected specifically for the purposes of this study; and the samples that were used would have otherwise been discarded. Samples were acquired and used in accordance with institutional guidelines.

Blood samples were used to investigate the hypothesis by use of 2 methods (4 experiments) to assess tonicity effects of various organic osmoles while excluding the effects of accompanying cations. First, a modified RBC osmotic fragility assay was used to compare tonicity effects of organic salts of ketoacids (sodium β-hydroxybutyrate and lithium acetoacetate) and lactate (sodium lactate and lithium lactate) with tonicity effects of inorganic salts (NaCl and LiCl). Second, a novel canine RBC assay was used to compare changes in RBC diameter after the addition of osmoles with known (glucose and urea) and unknown (β-hydroxybutyrate, acetoacetate, and lactate) effects.

MODIFIED OSMOTIC FRAGILITY ASSAY

Samples of RBCs were centrifuged at 264 X g for 10 minutes, followed by 3 serial washings with 300 mOsM NaCl (1 part RBCs to 3 parts NaCl). The final supernatant was discarded, and 100 µL of pelletted cells was resuspended in 500 µL of each test or control solution. Cells were incubated for 10 minutes at 22°C and then centrifuged at 435 X g for 3 minutes, and the pellet was discarded. As a marker for cell lysis, the free hemoglobin content of 100 µL of each solution was measured by spectral absorbance at 450 nm with a 96-well plate spectrophotometer. Samples were analyzed in duplicate, and values were expressed as percentage hemolysis, using RBCs incubated in double-distilled water to represent 100% hemolysis. Percentage hemolysis was determined by use of the following equation:

\[ \text{Percentage hemolysis} = \left( \frac{\text{OD}}{\text{OD}_{\text{ddH}_2\text{O}}} \right) \times 100\% \]

where OD is the optical density of the sample or control solution, and OD_{ddH_2O} is the optical density of double-distilled water and was used to represent 100% hemolysis.

First, relative tonicities of the ketoacid salts (sodium β-hydroxybutyrate and lithium acetoacetate) were compared with those of their respective chloride salts (NaCl and LiCl) by assessing their ability to protect RBCs from hypotonic-induced hemolysis. A series of sucrose solutions were prepared in 50mM increments (0 to 300mM; in triplicate). Then, 50mM sodium β-hydroxybutyrate or 50mM NaCl was added to each series. Results were determined for each series, and the unadulterated sucrose series was used as a control series. An identical procedure was performed with lithium acetoacetate and LiCl. The control series (sucrose) was replicated 4 times, and each sample series (sodium β-hydroxybutyrate, NaCl, lithium acetoacetate, and LiCl) was replicated 8 times; the mean for each data point was calculated. From these means, hemolysis protection curves for each series were generated by plotting the percentage hemolysis of each sample or control series against the sucrose concentration of each solution. For each curve, the H50 was visually determined; the H50 was compared between sodium β-hydroxybutyrate and NaCl, between lithium acetoacetate and LiCl, and between each sample series and the control (sucrose) solution. An H50 lower than that of the control series indicated an osmoprotective (tonicity) effect of the added (nonsucrose) solutes.

Next, the H50s of the control solution, NaCl, and sodium β-hydroxybutyrate were compared to assess whether the β-hydroxybutyrate anion exerts a tonicity effect independent of the sodium cation. The number of nonsucrose effective osmoles added to the NaCl-containing series, as determined by results for the osmotic fragility assay, was calculated by subtracting the H50 of the NaCl solution from the H50 of the control solution. This difference was divided by the nonsucrose effective osmoles known to be added to each solution (ie, 50mM NaCl = 100 mOsM) to establish a correction factor for the assay by use of the following equation:
Assay correction factor = (H50\text{control} - H50\text{NaCl})/100 \text{ mOsM}

The difference between the H50 of the control series and the H50 of the sodium β-hydroxybutyrate-containing series was divided by the assay correction factor to determine the number of nonsucrose effective osmoles that 50mM sodium β-hydroxybutyrate added to the solution. It was assumed that if β-hydroxybutyrate were an effective osmole, then sodium β-hydroxybutyrate would contribute 2 effective OsM/mole of salt (50mM sodium β-hydroxybutyrate = 100 mOsM), which represented the toxicity contribution of both sodium and β-hydroxybutyrate. However, if β-hydroxybutyrate were an ineffective osmole, then sodium β-hydroxybutyrate would contribute only 1 effective OsM/mole of salt (50mM sodium β-hydroxybutyrate = 50 mOsM), which represented only the toxicity contribution of sodium. A similar analysis was performed for lithium acetooacetate and LiCl. To assess the toxicity effect of lactate, these experiments were repeated by comparing the H50 of sodium lactate against that of NaCl and the H50 of lithium lactate against that of LiCl.

**CANINE RBC DIAMETER ASSAY**

Samples of RBCs were prepared by washing in a dilution with 0.01M PBS solution (1 part RBCs to 5 parts PBS solution). The cells were mixed and centrifuged at 202 X g for 5 minutes, and the supernatant was discarded. The wash step was repeated, and the cell pellet was then resuspended in a volume of 300 mOsM NaCl (1 part RBCs to 1 part NaCl). An aliquot of suspended RBCs was diluted in 300 mOsM NaCl (1 part suspended RBCs to 20 parts NaCl) and allowed to acclimate for at least 10 minutes at 22°C.

An aliquot (1 µL) of acclimated RBCs was added to 500 µL of each control or test solution and incubated for 30 minutes at 22°C. The RBC diameters in each solution were measured with a handheld automated cell counting device. After gentle mixing, a 100-µL sample containing RBCs was aspirated into a disposable 40-µm sensor attached to the cell counting device; at least 2.02 X 10^7 RBCs were counted with the cell counter for each analysis. Cell counts stratified by cell diameter in the form of a histogram were automatically generated by the cell counting device. Data were downloaded and imported into the cell counting device software, and the histograms were gated. Gating was performed manually by selecting the first histogram column from both ends of the range that was taller than it was wide (Figure 1). Mean RBC diameter was then calculated by the software and exported to a spreadsheet for data analysis.

First, osmoles of known toxicity effect were assayed to define the behavior of effective and ineffective osmoles in the canine RBC diameter assay. A 50mM solution of a known effective osmole (glucose) was prepared in 300 mOsM NaCl. Similarly, a 50mM solution of a known ineffective osmole (urea) was prepared in 300 mOsM NaCl. The RBC diameters for these solutions, as measured by the canine RBC assay, were compared against the RBC diameters for a 300 mOsM NaCl control solution.

Next, the toxicity effects of β-hydroxybutyrate, acetooacetate, and lactate were examined. The effects on RBC diameter of 300 mOsM solutions of sodium β-hydroxybutyrate and sodium lactate were compared with the effects of a 300 mOsM NaCl control solution. Because sodium acetooacetate was not readily available, acetooacetate was tested by use of the lithium salt. Solutions of 300 mOsM lithium acetooacetate and lithium lactate were prepared, and results for those solutions were compared against results for a 300 mOsM LiCl control solution. All experiments were performed in 15 to 18 replicates.

**STATISTICAL ANALYSIS**

The RBC diameter for all solutions was expressed as mean ± SD. The Wilcoxon rank test was used to compare RBC diameter between groups. Values of P ≤ 0.05 were considered significant.

**Results**

**MODIFIED OSMOTIC FRAGILITY ASSAY**

Hemolysis protection curves for sodium β-hydroxybutyrate and NaCl were plotted (Figure 2). All curves revealed a clear progression from 0% hemolysis (300mM sucrose) to 100% hemolysis (0mM sucrose). The H50 for both sodium β-hydroxybutyrate (78mM sucrose) and NaCl (31mM sucrose) was lower than that for the sucrose control solution (130mM sucrose); however, the H50 for NaCl was lower than that for sodium β-hydroxybutyrate. Similar relationships were evident between the H50 for lithium acetooacetate (87mM sucrose), LiCl (31mM sucrose), and the control solution (130mM sucrose).

The correction factor for the osmotic fragility assay with sodium salts was determined to be 0.99.
Therefore, the corrected nonsucrose effective osmoles from 50mM sodium β-hydroxybutyrate added to a solution was calculated as 51 mOsM ([130mM – 78mM]/0.99). This was almost exactly the effective osmoles expected to be added in accordance with the assumption that sodium β-hydroxybutyrate contributed only 1 effective OsM/mole of salt (50 mOsM). The correction factor for the osmotic fragility assay with lithium salts was also 0.99. Therefore, the corrected nonsucrose effective osmoles from 50mM lithium acetoacetate added to a solution was calculated as 42 mOsM ([130mM – 87mM]/0.99). This was closest to the effective osmoles expected to be added in accordance with the assumption that lithium acetoacetate contributed only 1 effective OsM/mole of salt (50 mOsM), as opposed to the value for the assumption that lithium acetoacetate contributed 2 effective OsM/mole of salt (100 mOsM).

Hemolysis protection curves for sodium lactate and NaCl were plotted (Figure 3). All curves revealed a clear progression from 0% hemolysis (300mM sucrose) to 100% hemolysis (0mM sucrose). The H50 for both sodium lactate and NaCl was 80mM sucrose and lower than that for the sucrose control solution (185mM sucrose). The H50 was similar for lithium lactate (75mM sucrose) and LiCl (80mM sucrose), and both were lower than that for the sucrose control solution (185mM).

The correction factor for the osmotic fragility assay with sodium salts was determined to be 1.05. Therefore, the corrected nonsucrose effective osmoles from 50mM sodium lactate added to a solution was calculated as 100 mOsM ([185mM – 80mM]/1.05). This was exactly the effective osmoles expected to be added in accordance with the assumption that lithium acetoacetate contributed 2 effective OsM/mole of salt (100 mOsM).

Hemolysis protection curves for sodium lactate and NaCl were plotted (Figure 3). All curves revealed a clear progression from 0% hemolysis (300mM sucrose) to 100% hemolysis (0mM sucrose). The H50 for both sodium lactate and NaCl was 80mM sucrose and lower than that for the sucrose control solution (185mM sucrose). The H50 was similar for lithium lactate (75mM sucrose) and LiCl (80mM sucrose), and both were lower than that for the sucrose control solution (185mM).
assumption that sodium lactate contributed 2 effective OsM/mole of salt (100 mOsM). The correction factor for the osmotic fragility assay with lithium salts was 1.10. Therefore, the corrected nonsucrose effective osmoles from 50mM lithium lactate added to a solution was calculated as 95 mOsM ([185 – 80mM]/1.10). This was closest to the effective osmoles expected to be added in accordance with the assumption that lithium lactate contributed 2 effective OsM/mole of salt (100 mOsM), as opposed to the value for the assumption that lithium lactate contributed only 1 effective OsM/mole of salt (50 mOsM).

**CANINE RBC DIAMETER ASSAY**

Results of experiments for osmoles of known tonicity effect were plotted (Figure 4). Mean ± SD RBC diameter in the glucose-containing solution (4.39 ± 0.067 µm) was smaller than, but not significantly (P = 0.56) different from, the mean RBC diameter in the NaCl control solution (4.40 ± 0.076 µm). However, the mean RBC diameter in the urea-containing solution (4.50 ± 0.088 µm) was significantly (P = 0.014) larger, compared with the mean RBC diameter in the NaCl control solution.

Results of experiments for osmoles of unknown tonicity effect (β-hydroxybutyrate, acetoacetate, and lactate) were plotted (Figure 5). Mean ± SD RBC diameter in the sodium β-hydroxybutyrate solution (5.43 ± 0.394 µm) was significantly (P < 0.001) larger, compared with the mean RBC diameter in the NaCl control solution (4.52 ± 0.190 µm). In contrast, mean RBC diameter in the sodium lactate solution (4.08 ± 0.122 µm) was significantly (P < 0.001) smaller, compared with the mean RBC diameter in the NaCl control solution. Mean RBC diameter was significantly larger in both the lithium acetoacetate solution (5.09 ± 0.194 µm; P < 0.001) and lithium lactate solution (4.33 ± 0.335 µm; P = 0.003), compared with the mean RBC diameter in the LiCl control solution (3.97 ± 0.239 µm).

**Discussion**

Analysis of results of the present study supported the hypothesis that the ketoacids β-hydroxybutyrate and acetoacetate acted as ineffective osmoles and therefore did not exert a tonicity effect. When tested by use of the modified osmotic fragility assay, both the NaCl and sodium β-hydroxybutyrate (NaBHB) and sodium lactate was significantly (P < 0.001) different from that for the NaCl control solution. In panel B, notice that mean RBC diameter was significantly greater for lithium lactate (P = 0.003) and lithium acetoacetate (LiAA; *P < 0.001) than that for the LiCl control solution.
and reduces the osmotic gradient across the RBC membrane, which decreases the likelihood of lysis. \(^1\) Comparison of the H50 for the NaCl versus sodium \(\beta\)-hydroxybutyrate and LiCl versus lithium acetoacetate series permitted the specific effect of an anion (\(\beta\)-hydroxybutyrate and acetoacetate) to be determined. Addition of 50mM NaCl or LiCl increased tonicity by 100 mOsm because these salts dissociate completely in aqueous solution. Similar to results for all salts, sodium \(\beta\)-hydroxybutyrate and lithium acetoacetate also dissociate completely in aqueous solution. If \(\beta\)-hydroxybutyrate were an effective osmole, 50mM sodium \(\beta\)-hydroxybutyrate would increase tonicity by 100 mOsm, but if \(\beta\)-hydroxybutyrate behaved as an ineffective osmole, 50mM sodium \(\beta\)-hydroxybutyrate would only increase tonicity by 50 mOsm; the same was true for lithium acetoacetate solutions. In both series, the osmoprotective effect of the ketoacid salt was less than that of the corresponding chloride salt, which suggested that the effective osmoles contributed by each ketoacid salt were less than those contributed by the corresponding chloride salt. Calculation of a correction factor allowed for the tonicity effects of ketoacids to be distinguished from those of the sodium and lithium cations. The concentrations of effective osmoles from the 50mM sodium \(\beta\)-hydroxybutyrate and 50mM lithium acetoacetate were 51 and 42 mOsm, respectively, which were both approximately 50 mOsm, as expected if \(\beta\)-hydroxybutyrate and acetoacetate were ineffective osmoles. Thus, analysis of results of the modified fragility assay supported the conclusion that \(\beta\)-hydroxybutyrate and acetoacetate acted as ineffective osmoles and did not exert tonicity effects.

In contrast to the ketoacids, lactate appeared to act as an effective osmole in the osmotic fragility assay. With sodium or lithium as a cation, the difference in H50 between lactate and chloride was minimal; 50mM of both the chloride salt and lactate salt contributed approximately 100 mOsm to the osmoprotective effect of the solutions, as would be expected when both the cation and anion are effective osmoles. Interestingly, the H50s of NaCl, LiCl, and the sucrose control solution differed between the ketoacid and lactate experiments. Different RBC samples were used in the ketoacid and lactate experiments; therefore, it is possible that differences in intrinsic osmotic fragilities of these samples contributed to the discrepancy. However, the correction factor and comparison of the ketoacids and lactate relative to chloride were used to mathematically account for these differences.

In a second set of experiments, effects of urea and glucose were assessed with a novel assay to determine changes in RBC diameter. The RBC diameter was increased significantly by the addition of urea, an ineffective osmole. The increase in RBC diameter was expected because RBCs are highly permeable to urea.\(^{16}\) Influx of urea is accompanied by water that causes swelling of the cells and increases RBC diameter. Exposure to glucose, a known effective osmole, was expected to decrease RBC diameter through an opposite mechanism that encouraged water efflux and decreased cell size.\(^1\) In experiments with glucose, the RBC diameter decreased, albeit not significantly, relative to that for the control solution. The reason for the reduced effect of glucose in these test conditions was not clear but could have been caused by partial permeability of the RBC membrane to glucose. In particular, it has been reported that glucose transport into canine RBCs is higher than in many other species.\(^{17}\) It is possible that a high intracellular glucose concentration opposed the transcellular gradient induced by glucose incubation, which reduced the tonicity effect of the extracellular glucose. Although traditional physiology texts classify glucose as an effective osmole, some authors dispute this classification and argue that the tonicity effect of glucose is tissue and insulin dependent, given that both affect glucose transporter expression.\(^{1,2}\) In fact, the tonicity effect of almost all solutes is determined by membrane transporter expression, which changes on the basis of the cell milieu. Thus, it could be argued that effectiveness of a solute should be treated as a relative and malleable property, rather than given a binary classification. However, the urea and glucose experiments yielded distinct responses in RBC diameter. Thus, for the purposes of the present study as well as for simplicity, the behaviors of urea and glucose were considered the standard for ineffective and effective osmoles, respectively.

Incubation with sodium \(\beta\)-hydroxybutyrate and lithium acetoacetate significantly increased RBC diameter, compared with RBC diameter for the control solutions of NaCl and LiCl. Changes in RBC diameter caused by solutions of ketoacids were consistent with changes caused by incubation with urea, which suggested these substances act as ineffective osmoles. This is in accordance with the hypothesis and results from the modified fragility assay. However, changes in RBC diameter caused by lactate were discordant. Sodium lactate caused a decrease in RBC diameter relative to that for the NaCl control solution, which was a response expected for an effective osmole and consistent with the osmotic fragility results for lactate. Conversely, lithium lactate caused an increase in RBC diameter, compared with that of the LiCl control solution, which was consistent with action as an ineffective osmole. The reason lactate appeared to function differently when paired with different cations was not clear. The lithium concentration in these in vitro solutions exceeded that typical for RBCs and may have affected cell membrane function. For example, lithium could alter the function of the monocarboxylate transporter, which facilitates entry of lactate into cells.\(^{18}\) If this were the case, then discordant osmotic fragility results between sodium lactate and lithium lactate might also be expected; however, the total lithium concentration used in the RBC diameter assay was much higher (150mM) than that used in the osmotic fragility assay (50mM), which could create a differential effect. The different lithium concentrations may also explain the reason that the RBC diameter of NaCl (4.52 μm) was higher than that of LiCl (3.97 μm) but the H50s were the same (80mM).
cross). The monocarboxylate transporters also mediate transport of ketoacids, so the toxicity effect of acetoc-
ate, which was measured as a lithium salt, might also be affected by a supranormal lithium concentration.18
Unfortunately, acetocacetate is not readily available as a sodium salt, so comparisons between the sodium and lithium salts of acetocacetate could not be determined.

A limitation of the present study was that only RBCs were evaluated. Toxicity effects of solutes may differ by tissue type and disease state, so the effects of ketoacids and lactate on RBCs may not represent the response for all cell types. For practical reasons (ie, ease of simple collection and handling) RBCs were used, but RBCs also serve as a classic model for biological osmo-
lality studies.19,20 However, other cell lines could be con-
sidered in future experiments to assess effects on target tissues that are particularly susceptible to toxicity in-
jury, such as neural tissues.5 The test solutions used in these experiments were simple isotonic or hypertonic aqueous solutions, which were needed to isolate the ef-
fects of individual osmole. Ideally, RBC diameter would have been measured in solutions that approximated the composition of serum or interstitial fluid. Serum was not used for RBC diameter experiments because of technical limitations of the cell counting device used to determine RBC diameter. Similarly, technical limitations also precluded evaluation of RBCs in hypotonic solu-
tions. The RBC diameters yielded by the cell counting device were smaller than that reported for typical ca-
nine RBCs (7 µm).21 This may reflect a technical charac-
teristic of the device used to measure RBC size or may reflect changes caused by the storage and processing of the cells. A methodological difference appears the most likely cause for the discrepancy between previously re-
ported RBC diameters and those in the present study because the mean corpuscular volume of RBCs stored at 4°C can increase over time.22 Regardless, all cells used were directly compared with cells that underwent identical in vitro processing and measurement; there-
fore, conclusions regarding toxicity effects of solutes were likely valid but extrapolations to in vivo systems should be made with caution.

In the biological models of the present study, β-hydroxybutyrate and acetocacetate acted as ineffective osmole,
ways, lactate predominately acted as an ef-
flective osmole. On the basis of these findings, it is un-
likely that β-hydroxybutyrate and acetocacetate would contribute to toxicity in clinical syndromes, such as di-
abetes mellitus or ketoacidosis; however, it is possible that severe lactic acidosis could affect toxicity of the milieu.

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Footnotes
a. SpectraMax190, Molecular Devices, Sunnyvale, Calif.
b. Sigma-Aldrich, St Louis, Mo.
c. Standard solution, 300 mOsm/kg H2O, Precision Systems, Na
tick, Mass.
d. Scepter, version 2.0, EMD Millipore, Hayward, Calif.
e. Scepter Software Pro, version 2.0, EMD Millipore, Hayward, Calif.
f. Excel, Microsoft, Redmond, Wash.

References
1. Wellman ML, DiBartola SP, Kohn CW. Applied physiology of body fluids in dogs and cats. In: DiBartola SP ed. Fluid, elec-
3. Garner MM, Burg MB. Macromolecular crowding and confi-
7. Schermerhorn T, Barr SC. Relationships between glucose, so-
13. Cox K, Cocchi MJ, Salciccioli JD, et al. Prevalence and signifi-
16. Liu L, Lei T, Bankir L, et al. Erythrocyte permeability to urea and water: comparative study in rodents, ruminants, carnivores, hu-
17. Arai I, Washizu T, Sagara M, et al. β-glucose transport and glyco-
20. Hendry EB. Osmolarity of human serum and of chemical solu-
22. Furlanello T, Tasca S, Caldin M, et al. Artifactual changes in ca-