Strangulating volvulus of the large colon is one of the most severe forms of colic and can account for 11% to 27% of horses requiring surgical correction of colic in referral hospitals. Survival after surgery for correction of large colon volvulus is dependent on the degree of ischemic injury to the colon and severity of the systemic response. The survival rate without surgical resection has been reported to be as low as 34.7%, but resection improved the survival rate of affected horses in 2 studies (57.7% and 74%, respectively). In most horses that undergo colon resection, some ischemic tissue remains. Thus, survival in these horses can be correlated with loss of the epithelial barrier in the remnant portion, which allows transmucosal leakage of endotoxins, bacterial chemotactic peptides, and bacteria. Therefore, rapid repair of the ischemic-injured epithelium is crucial to recovery of a horse after large colon volvulus but could be impaired by reperfusion injury.

The biochemical pathway responsible for reperfusion injury starts with accumulation of products that build up during ischemia and ROS generated on reperfusion. Neutrophil infiltration is a time-dependent process that occurs mostly during the reperfusion period, and activation of these cells results in degranulation and additional release of inflammatory mediators, such as cytokines, ROS, proteases, and other regulatory proteins. Although ischemia is a major pathophysiological event in strangulating volvulus of the large colon, the role of reperfusion damage is also significant in determining the outcome of affected horses. In vitro and in vivo responses of mucosa from the large colon of horses to ischemia and reperfusion

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**Objective**—To induce ischemia and reperfusion injury in the large colon mucosa of horses in vivo and evaluate the recovery and effects of components of an organ transplant solution on mucosal recovery in vitro.

**Animals**—6 healthy horses.

**Procedures**—Horses were anesthetized, and ischemia was induced for 60 minutes in the pelvic flexure, which was followed by reperfusion for 240 minutes. Ischemic (n = 4 horses), reperfused (6), and adjacent control (6) colonic mucosae were isolated for in vitro testing and histologic examinations. Tissues were mounted in Ussing chambers with plain Krebs Ringer bicarbonate (KRB), KRB with N-acetylcysteine (NAC), or KRB with a modified organ transplant solution (MOTS). Transepithelial electrical resistance (TER) and mannitol flux were used to assess mucosal integrity. Data were analyzed by use of ANOVA and Kruskal-Wallis tests.

**Results**—The TER in reperfused tissues was similar to the TER in control tissues and greater than the TER in ischemic tissues, which was consistent with morphological evidence of recovery in reperfused tissues. Mannitol flux was greater in ischemic tissues than in reperfused tissues. The TER and mannitol flux were not significantly affected by incubation of mucosa with NAC or MOTS.

**Conclusions and Clinical Relevance**—Ischemia induced during the brief period allowed rapid mucosal repair and complete recovery of tissue barrier properties during reperfusion. Therefore, reperfusion injury was not observed for this method of ischemic damage in equine colonic mucosa. (Am J Vet Res 2011;72:982–989)
A olopathic feature of large colon volvulus, exacerbation of the ischemic injury during reperfusion has been detected during experimentally induced low-flow ischemia in equine colon. However, the importance of reperfusion injury in the equine colon remains controversial.

Studies on reperfusion injury primarily have been directed at detecting an exacerbation of ischemic damage during reperfusion, with or without protection against that damage via various interventions. A novel OPS used in human kidney transplant patients was able to preserve viability in an isolated segment of equine colon in the absence of oxygen and blood. The solution represents a multimodal approach to treating reperfusion injury but is expensive. However, 3 of its components are inexpensive and warrant study separately as treatments for reperfusion injury. The amino acid l-arginine is an indirect precursor of nitric oxide and therefore has the potential to hasten tissue healing and modulate the inflammatory response. The amino acid l-glutamine can influence several metabolic pathways during intestinal stress and was protective in an in vitro evaluation of chemical damage to equine colonic mucosa. Metabolism of arginine and glutamine is closely related, and these amino acids could result in an additive effect on several biochemical pathways. Acetyl cysteine can protect mucosa through its antioxidant properties and by providing sulfhydryl groups required for replenishment of glutathione, which is an important intracellular antioxidant. In equine colonic mucosa, acetyl cysteine can prevent mucosal damage and reduce eosinophil migration after chemical damage in vitro.

Our hypothesis was that experimentally induced ischemia of short duration would cause mild to moderate reversible injury to equine colonic mucosa and that this injury would be exacerbated during reperfusion. The purpose of the study reported here was to evaluate mucosal damage of the equine colon caused by ischemia and reperfusion, as determined on the basis of histomorphometric changes; determine TER and permeability to mannitol in tissues subjected to ischemia and reperfusion, compared with results for control tissues and tissues subjected to ischemia only; and evaluate in vitro effects of components of an OPS on functional and morphological measurements of mucosal recovery.

**Materials and Methods**

**Horses**—Six healthy adult horses (3 Thoroughbreds, 2 Quarter horses, and 1 warmblood) were included in the study. Horses were 11 to 26 years old (mean, 16 years) and weighed 440 to 660 kg (mean, 548 kg). Inclusion criteria were that the horses were in good health, were free of gastrointestinal tract disease, and required euthanasia for conditions that rendered them unsuitable for use. Horses were housed on pasture with grass hay and water provided ad libitum and did not receive any medications for the 2-week period preceding the study. The Institutional Animal Care and Use Committee of the University of Florida approved the study protocol.

**Surgical procedure**—Horses were sedated with xylazine (1 mg/kg, IV). A 14-gauge, 13.3-cm polytetrafluoroethylene catheter was inserted into each of the jugular veins; the left jugular vein was used for administration of anesthetic drugs and isotonic fluids, and the right jugular vein was used for collection of blood samples. Anesthesia was induced with ketamine (2.2 mg/kg, IV) and diazepam (0.1 mg/kg, IV). Orotracheal intubation was performed and horses were positioned in dorsal recumbency. Anesthesia was maintained with isoflurane (1% to 3%) in oxygen via mechanical ventilation. Isotonic polyionic fluids were continuously infused at a rate of 2.5 to 5 mL/kg/h. Mean arterial blood pressure was monitored through a 20-gauge, 5.1-cm polytetrafluoroethylene catheter in the facial artery and was maintained at ≥ 60 mm Hg. Monitoring performed during anesthesia included electrocardiography, arterial blood gas analysis, capnography, and direct blood pressure measurement.

The ventral abdomen of each horse was prepared and draped for aseptic surgery, and ventral midline celiotomy was performed. The large colon was exteriorized and positioned on a sterile drape on the ventral abdomen. To induce ischemia, a 40-cm segment of colon at the pelvic flexure was subjected to transmural compression via intestinal clamps placed at each end, and venous and arterial occlusion was achieved with umbilical tape ligatures. After induction of ischemia, the colon, colonic vasculature, and associated mesentery were surgically divided at the pelvic flexure so that 2 segments of colon comparable in size (dorsal and ventral) did not communicate. The colon was then replaced in the abdomen, and the abdominal incision was closed temporarily with towel clamps. After a 1-hour period of ischemia, the colon was again exteriorized and 1 of the 2 ischemic segments (ischemic tissues) was resected for histologic evaluations and in vitro experiments in Ussing chambers. Ischemic tissues were not harvested for Ussing chamber experiments from the first 2 horses; thus, tissues from only 4 horses were used for this phase of the study. At the same time, the clamps and ligatures were removed from the other segment of ischemic colon; this segment was replaced in the abdomen to allow resumption of blood flow (reperfusion) for 4 hours (horses remained anesthetized during reperfusion). Small (1- to 2-cm) mucosal biopsy specimens were collected before ischemia, after a 1-hour period of ischemia (time 0 for ischemic tissues), and after reperfusion for 1, 2, and 4 hours. After reperfusion for 4 hours, the reperfused segment of colon (reperfused tissues) and an adjacent nonischemic-nonreperfused segment of colon (control tissues) were removed (time 0 for control and reperfused tissues) for histologic evaluation and in vitro experiments in Ussing chambers. After the control and reperfused tissues were harvested, the anesthetized horses were euthanized with an overdose of sodium pentobarbital (88 mg/kg, IV).

**Ussing chamber experiments**—A solution of KRB (112 mM NaCl, 25 mM NaHCO₃, 10 mM glucose, 5 mM KCl, 3 mM sodium acetate, 3 mM sodium butyrate, 2.5 mM CaCl₂, 1.2 mM MgSO₄, 1.2 mM KH₂PO₄, and 0.01 mM mannitol [pH, 7.4]) was prepared. After removal, full-thickness tissue sections from ischemic, reperfused, and control colon were immediately placed in cold (4°C) KRB solution and transported to our labo-
The timing of sample collection and mounting of tissues in Ussing chambers for the in vitro experiments was such that experiments were completed on the ischemic tissues before the same experiments were performed on control and reperfused tissues. Mucosal sheets from each colon segment (control, ischemic, and reperfused tissues) were removed and mounted in Ussing chambers as described elsewhere. Three treatment solutions were used to incubate the mucosal tissues: KRB alone; KRB with 70 mg of NAC/L (KRB-NAC treatment); and KRB with 5 mg of l-arginine/L, 70 mg of NAC/L, and 10 mg of l-glutamine/L (KRB-MOTS treatment). Each tissue incubation was performed in duplicate.

The short-circuit current was recorded in each chamber by use of voltmeter clamps through silver-silver chloride electrodes connected to 4% agar bridges in KRB solution. Junction potentials of electrodes and fluid resistance were measured before mounting of the tissues to allow continuous correction for any effects that these factors might have on the low potential differences generated by the tissues. When the tissues were mounted in the chambers, the voltage clamp could then automatically correct for junction potentials of electrodes and fluid resistance and reduce their effects on the recordings. Throughout incubation, the short-circuit current was continuously applied to the tissues, except for a brief period at 15-minute intervals when the potential difference of the tissue was measured. The TER was calculated by use of Ohm’s law, whereby resistance is equal to the potential difference of the tissue divided by the short-circuit current. Resistance was used as a measure of integrity of colonic mucosa and permeability of the paracellular pathway to ions.

The unidirectional flux of tritium-labeled mannitol from the mucosal to the serosal solution was an additional measure of colonic mucosa permeability. For scintillation counting, fluid samples were collected from both sides (mucosal and serosal) at 45, 75, 105, and 240 minutes after addition of radiolabeled mannitol to the mucosal side. Transmucosal flux of mannitol was expressed as the percentage of the initial scintillation counts detected on the mucosal side of the tissue that was detected on the serosal side of the tissue at the 4 time points. The interval from tissue collection to mounting of the tissue in the first chamber was approximately 10 minutes and to first recording of TER and addition of tritium-labeled mannitol was approximately 40 minutes. Total incubation time for each tissue in the Ussing chamber was sufficient to record TER and mucosal flux for 240 minutes. After Ussing chamber experiments were completed, the tissues were removed and placed in neutral-buffered 10% formalin for histologic evaluation and histomorphometric measurements.

Histologic evaluation and histomorphometric measurements—Biopsy specimens of colonic mucosa obtained in vivo before induction of ischemia (time 0), after a 1-hour period of ischemia, and after reperfusion for 1, 2, and 4 hours and in vitro at the end of Ussing chamber experiments were fixed in neutral-buffered 10% formalin. Tissues were embedded in paraffin, cut into 5-µm-thick sections, placed on silane-coated glass slides, and stained with H&E for examination via light microscopy. For histomorphometric assessment of mucosal damage via light microscopy, a computer-based imaging-analysis program was used and 3 fields from each tissue were examined as described elsewhere. One investigator (AG) was unaware of the treatment group for each biopsy specimen, performed all histologic evaluations.

The length of denuded epithelium was measured and expressed as a percentage of the total mucosal length in the section. The epithelium was defined as lifted when > 5 epithelial cells were separated from the basement membrane but still attached to adjacent cells. The length of epithelium affected by lifting was expressed as a percentage of the total surface length. Detached cells were defined as cells that appeared morphologically normal but were separated from the basement membrane in groups of ≥ 5 cells and completely detached from adjacent epithelium. The length of detached epithelium was measured and expressed as a percentage of the total surface length of the mucosa. Restituted cells were defined as cells that were flattened in appearance but had intact attachments to the basement membrane and to adjacent cells. The length of epithelium consisting of restituted epithelial cells was measured and expressed as a percentage of the total surface length of the mucosa. The number of sloughed cells was counted for each field; the mean number of sloughed cells/0.1 mm of surface length was calculated. Sloughed cells were not counted in tissue samples incubated in the Ussing chambers because there were no sloughed cells present or the sloughed cells were of undetermined origin.

Statistical analysis—The tissues obtained from each horse yielded 2 sets of observations for each experimental condition in the Ussing chambers. The mean of the data was calculated and expressed as least squares mean ± SEM, and a statistical software program was used for analysis. Repeated-measures ANOVA was performed on the TER for each of the 3 tissue groups. Whenever there was a significant result of an F test for treatment, time, or the treatment-by-time interaction, appropriate Bonferroni-adjusted P values were used for each family of comparisons. To determine the effect of ischemia and reperfusion on tritium-labeled mannitol flux after incubation in the Ussing chambers for 240 minutes, a 1-way ANOVA was performed. Statistical analysis of histologic data was performed by use of a Kruskal-Wallis test to determine significant differences in histomorphometric measurements between the treatment groups. Post hoc analysis was performed by use of the Mann-Whitney U test. Values of P < 0.05 were considered significant for all statistical analyses.

Results—Ussing chamber experiments—The TER was not significantly changed by the Ussing chamber treatments (KRB, KRB-MOTS, and KRB-NAC) within each of the 3 tissue groups (control tissues [n = 6 horses], ischemic tissues [4], and reperfused tissues [6]). The TER of the control tissues decreased significantly with time during the incubation period, whereas there was
Figure 1—Mean ± SEM values for TER of colonic mucosa obtained from anesthetized horses after ischemic conditions for 1 hour (ischemic tissue [solid line and black symbols]) or after ischemic conditions for 1 hour followed by reperfusion for 4 hours (reperfused tissue [dashed lines and black symbols]) or from adjacent control segments after ischemia-reperfusion (control tissue [solid line and white symbols]). Each time point represents the mean value for samples (evaluated in duplicate) obtained from 6 horses (control and reperfused tissues) or 4 horses (ischemic tissue). Time 0 = end of 1 hour of ischemia (ischemic tissues) or end of 4 hours of reperfusion (control and reperfused tissues). Immediately after collection, tissues were mounted in Ussing chambers and incubated with KRB (circles), KRB and MOTS (KRB-MOTS treatment [squares]), or KRB and NAC (KRB-NAC treatment [triangles]) for 4 hours. Values did not differ significantly (P ≥ 0.05) among treatments at any time within the control, ischemic, or reperfused tissue groups. Values for all reperfused tissues differed significantly (P < 0.05) from values for all ischemic tissues, except when comparing reperfused and ischemic tissues incubated in KRB-MOTS. *Within a time point, values for all control tissues (KRB, KRB-NAC, and KRB-MOTS) differed significantly (P < 0.05) from values for all ischemic tissues. †Within a time point, values for all reperfused tissues (KRB, KRB-NAC, and KRB-MOTS) differed significantly (P < 0.05) from values for all ischemic tissues. ‡Within a time point, values for all control tissues differed significantly (P < 0.05) from values for all ischemic tissues, except when comparing control tissues incubated in KRB or KRB-MOTS with values for ischemic tissues incubated in KRB-MOTS.

Figure 2—Mean ± SEM values for permeability of equine colonic mucosa to tritium-labeled mannitol for control, ischemic, and reperfused tissues after the tissues were incubated in an Ussing chamber for 240 minutes. Tissues were incubated in KRB (gray bars), KRB-MOTS (white bars), or KRB-NAC (black bars). Transmucosal flux of mannitol was expressed as the percentage of the initial scintillation counts detected on the mucosal side of the tissue that were detected on the serosal side of the tissue at 4 time points (ie, 45, 75, 105, and 240 minutes after addition of radiolabeled mannitol to the mucosal side). *Value differs significantly (P < 0.05) from the value for reperfused mucosa incubated in KRB.
There was a significant increase in the percentage of denuded epithelium for the ischemic tissues, compared with the percentage for the control and reperfused tissues. Compared with results for the control tissues, ischemic tissues had a significantly greater percentage of denuded epithelium and greater epithelial restitution, compared with results for the reperfused tissues; however, these values did not differ significantly. Results for the reperfused tissues were not significantly different from results for the control tissues with respect to any of the variables examined. There was no significant difference in the percentage of lifted epithelium or degenerated epithelium among any of the groups. There was no significant effect of Ussing chamber treatments (KRB, KRB-NAC, or KRB-MOTS) on the histomorphometric measurements; therefore, all statistical analyses were performed for the tissues incubated in KRB. When the in vivo mucosal biopsy specimens were compared with the in vitro biopsy specimens, the most notable difference was evidence of epithelial cell restitution in the ischemic tissues after incubation in the Ussing chambers for 240 minutes (Figure 3). Predictably, all tissues had a reduced intensity of staining in the lamina propria after incubation in the Ussing chambers for 240 minutes.

**Discussion**

In the present study of experimentally induced ischemia in the equine large colon, reperfusion did not exacerbate the ischemic damage but appeared to allow mucosal recovery, as determined on the basis of histologic and functional measurements of tissue integrity. The ischemia experimentally induced in the colon of horses in the study reported here caused mild injury that should have allowed some expression of reperfusion injury over the next 4 hours. A more severe ischemic injury may not have allowed a sufficient number of viable cells to survive and cause an obvious reperfusion injury. However, further studies are required to determine whether 1 hour is an optimum ischemic period to allow generation of ROS and the associated tissue damage in equine colonic mucosa.

In the original experimental design, responses of interest were to be examined in 2 separate groups of horses that would be assigned by use of a randomization procedure to an ischemia group and an ischemia-reperfusion group. However, it was evident after the procedure was performed on the first 2 horses in the ischemia-reperfusion group that the experimental procedures allowed sufficient time for investigators to mount the ischemic tissues in the Ussing chambers.
cubate the tissues, and then empty and rinse the chambers in time for the reperfused tissues to be mounted. In this manner, each horse could be used to provide tissues subjected to ischemia and to reperfusion, and the difference between the 2 tissues was sufficient such that only 4 sets of ischemic tissue were required to yield significant differences.

When the function of the mucosa was examined in Ussing chambers, the ischemic tissues performed significantly worse than did the control and reperfused tissues, as measured on the basis of TER. There was no significant difference between the TER for the control and reperfused tissues. Similarly, the reperfused tissues were less permeable to mannitol than were the ischemic tissues, which indicated some restoration of barrier function in the reperfused tissues.

Histomorphometric examination also revealed restitution in reperfused tissues with almost complete mucosal recovery after 4 hours in vivo. This early recovery of mucosal epithelium was mainly attributable to migration of surviving epithelial cells over the damaged surface and not by cell proliferation, which would require a considerably longer observation period in vivo. 

Complete arteriovenous occlusion for 1 hour was used in the present study to closely mimic large colon volvulus in horses. A period of 4 hours of reperfusion was chosen as a reasonable amount of time in which observations could be made with horses remaining anesthetized. In other studies, in which investigators used similar ischemic methods for the equine colon and jejunum, a reperfusion period of 18 hours had been used, which necessitated recovery of the horses from anesthesia. Because mucosal restitution and tightening of paracellular pathways are critical for recovery, the time period chosen for examination of the effects of reperfusion should be appropriate to evaluate these aspects of early repair. The findings in the study reported here also are consistent with findings in another study26 conducted by our research group in which we found that a more severe mucosal injury followed by reperfusion for 18 hours also allowed mucosal recovery, although to a lesser extent than in the present study. Although only 4 horses were used in the ischemic group in the present study, changes were identical to those reported for this duration of ischemia in tissues of the pelvic flexure of horses.27,28

Selected components (NAC and MOTS) of a novel OPS solution17 were added to Ussing chambers to determine whether these components could improve recovery in vitro.20,21 These components were chosen rather than the OPS solution described elsewhere21 because, if effective, they could be administered systemically in a clinical setting at low cost. Some benefit has been proposed for combining arginine and glutamine,19 and inclusion of acetylcysteine is warranted on the basis of salutary effects in horses29 and rats with chemically induced mucosal injury.14 There was no evidence of any improvement in recovery from mucosal injury in vitro for the conditions of the present study, although the changes induced by this reversible ischemia method would have seemed appropriate to test such an effect.

The MOTS components were applied directly to the mucosal and serosal surfaces to improve access to epithelial and inflammatory cells in the mucosa. This design could only be expected to detect an effect on epithelial cells and on neutrophils already present in the tissues in vitro, without an opportunity to evaluate

Table 1—Mean ± SEM histomorphometric values for control (n = 4 horses), ischemic (6), and reperfused (6) colonic tissues after incubation with KRB, KRB-MOTS, or KRB-NAC in Ussing chambers for 4 hours.

<table>
<thead>
<tr>
<th>Variable</th>
<th>Control</th>
<th>Ischemic</th>
<th>Reperfused</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mucosal height (µm)</td>
<td>401.59 ± 17.34</td>
<td>390.15 ± 16.74</td>
<td>406.61 ± 22.67</td>
</tr>
<tr>
<td>Epithelial height (µm)</td>
<td>27.23 ± 1.72</td>
<td>25.75 ± 0.94</td>
<td>27.47 ± 1.26</td>
</tr>
<tr>
<td>Epithelial width (µm)</td>
<td>4.84 ± 0.20</td>
<td>4.96 ± 0.13</td>
<td>5.02 ± 0.22</td>
</tr>
<tr>
<td>Denuded epithelium (%)</td>
<td>0.02 ± 0.02</td>
<td>0.05 ± 0.05</td>
<td>0.13 ± 0.13</td>
</tr>
<tr>
<td>Lifted epithelium (%)</td>
<td>0.00 ± 0</td>
<td>0.00 ± 0</td>
<td>0.00 ± 0</td>
</tr>
<tr>
<td>Degenerated epithelium (%)</td>
<td>0.00 ± 0</td>
<td>0.00 ± 0</td>
<td>0.00 ± 0</td>
</tr>
<tr>
<td>Restituted epithelium (%)</td>
<td>0.00 ± 0</td>
<td>0.00 ± 0</td>
<td>0.00 ± 0</td>
</tr>
<tr>
<td>Necrotic or sloughed cells/0.1 mm</td>
<td>0.11 ± 0.03</td>
<td>2.97 ± 0.96</td>
<td>2.20 ± 1.15</td>
</tr>
</tbody>
</table>

*Within a row, values with different superscript letters differ significantly (P < 0.05).

Table 2—Mean ± SEM histomorphometric values for control (n = 4 horses), ischemic (6), and reperfused (6) colonic tissues after incubation with KRB, KRB-MOTS, or KRB-NAC in Ussing chambers for 4 hours.

<table>
<thead>
<tr>
<th>Variable</th>
<th>Control</th>
<th>Ischemic</th>
<th>Reperfused</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mucosal height (µm)**</td>
<td>36.13 ± 1.86</td>
<td>25.64 ± 1.11</td>
<td>26.21 ± 1.58</td>
</tr>
<tr>
<td>Epithelial height (µm)**</td>
<td>3.72 ± 0.10</td>
<td>4.87 ± 0.13</td>
<td>4.62 ± 0.27</td>
</tr>
<tr>
<td>Lifted epithelium (%)</td>
<td>0.00 ± 0</td>
<td>0.97 ± 0.47</td>
<td>0.00 ± 0</td>
</tr>
<tr>
<td>Degenerated epithelium (%)</td>
<td>0.56 ± 0.22</td>
<td>0.78 ± 2.40</td>
<td>0.96 ± 0.45</td>
</tr>
<tr>
<td>Restituted epithelium (%)</td>
<td>0.00 ± 0</td>
<td>0.00 ± 0</td>
<td>10.30 ± 5.23</td>
</tr>
<tr>
<td>Necrotic or sloughed cells/0.1 mm</td>
<td>0.11 ± 0.03</td>
<td>2.97 ± 0.96</td>
<td>2.20 ± 1.15</td>
</tr>
</tbody>
</table>

*Within a row, values with different superscript letters differ significantly (P < 0.05).

See Table 1 for remainder of key.
any putative antioxidant effect on neutrophil infiltration. Therefore, further investigation into the potential clinical use of these components on the inflammatory response in reperfused mucosa might be better conducted in vivo. The rapid recovery of barrier function during reperfusion, as measured on the basis of TER, can be attributed to increased expression of tight junction proteins (occludin and ZO-1) during mucosal recovery. Application of prostaglandin E, to ischemia-injured ileal mucosa stimulates increases in TER initiated through chloride secretion in restituting epithelium, which suggests that cyclooxygenase enzymes may play an important role in intestinal protection and recovery. The marked upregulation of cyclooxygenase-2 expression in the epithelial and crypt cells of the equine colon after ischemia may produce the production of prostaglandin E, and other prostanoids during reperfusion, which could enhance epithelial recovery and restitution of barrier function.

Anesthetic preconditioning via administration of volatile anesthetics, such as isoflurane, before ischemia can diminish the severity of ischemia-reperfusion injury in the brain, heart, kidneys, liver, and lungs in humans and other animals. To our knowledge, the use of preconditioning with volatile anesthetics has not been found to reduce ischemia-reperfusion injury in the intestines. In the present study, isoflurane was used to anesthetize the horses for ischemia and reperfusion periods and may have reduced the severity of the ischemic injury or attenuated the degree of reperfusion injury through its anti-inflammatory effects. However, the clinical relevance of such preconditioning may be questionable because large colon volvulus is typically corrected in horses that are anesthetized by use of an anesthetic regimen similar to that used in the present study. Therefore, all horses undergoing surgery for large colon volvulus will have some anesthetic preconditioning before reperfusion.

The TER in reperfused tissues in the present study was sustained throughout the in vitro phase at values close to the maximum values for the control tissues, whereas TER values in control tissues decreased toward the end of incubation (Figure 1). Tissues subjected to ischemic conditions for 1 hour had lower TERs in the end of incubation (Figure 1). Tissues subjected to ischemic conditions for 1 hour had lower TERs in the end of incubation (Figure 1). Tissues subjected to ischemic conditions for 1 hour had lower TERs in the end of incubation (Figure 1).

Ischemic equine jejunum has significantly higher values throughout incubation in vitro than does the control jejunum. These findings can be explained by populations of tight junctions throughout the jejunum that are more permeable in control conditions but that become recruited into the intense and widespread process of tight junction closure after injury. The difference in expression of this process between jejunum and colon could be explained by the importance of a permeable paracellular pathway for nutrient absorption in jejunal epithelium. This amount of permeability could be lacking in the colon, which has a limited capacity to absorb nutrients and a greater need to have a tight epithelial barrier against noxious luminal contents.

The lack of reperfusion injury in the present study is consistent with results of other studies in horses and raises concerns about the relevance of this process in the equine intestines. However, results reported for a low-flow method of ischemia in the equine colon revealed a marked influx of neutrophils associated with continued mucosal degeneration during reperfusion. In clinically affected horses with large colon volvulus, the relative roles of low-flow ischemia and complete ischemia are unknown, so the most suitable method for investigation of this disease has not been established. However, the nature of the lesions would suggest that almost complete cessation of blood flow, as was induced in the present study, should be the most likely vascular change.

In equine colonic mucosa subjected to periods of ischemia similar to that used in the present study, but with shorter and longer intervals of reperfusion, rapid and intense neutrophil influx was evident throughout reperfusion. Although neutrophil influx was not measured in the present study, marked increases in neutrophil numbers were observed after reperfusion for 4 hours, a time point at which tissues had recovered from ischemic damage. For most methods of intestinal reperfusion injury, influx of neutrophils of such magnitude would be expected to exacerbate the ischemic injury. The role of neutrophils in reperfusion injury is further complicated by evidence that reperfusion-induced mucosal damage in equine jejunum can be independent of neutrophils.

We concluded for the experimental methods of the study reported here that complete ischemia of equine colonic mucosa for 1 hour followed by reperfusion for 4 hours did not result in functional or morphological evidence of reperfusion injury. Therefore, in horses that undergo surgery for large colon volvulus, efforts to hasten mucosal recovery probably should have precedence over aggressive efforts to treat reperfusion injury.

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c. Image Pro Express, version 5.0, Media Cybernetics Inc, Bethesda, Md.
d. SAS, version 8, SAS Institute Inc, Cary, NC.


