Effects of treatment with polysulfated glycosaminoglycan on serum cartilage oligomeric matrix protein and C-reactive protein concentrations, serum matrix metalloproteinase-2 and -9 activities, and lameness in dogs with osteoarthritis

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Objective—To investigate the effects of polysulfated glycosaminoglycan (PSGAG) treatment on serum cartilage oligomeric matrix protein (COMP) concentration, matrix metalloproteinase-2 (MMP-2) and -9 (MMP-9) activities, C-reactive protein (CRP) concentration, and lameness scores in dogs with osteoarthritis.

Animals—16 dogs with osteoarthritis and 5 clinically normal dogs.

Procedures—Dogs with osteoarthritis had a history of chronic lameness, and osteophytes were observed on radiographic evaluation of the affected joint. Polysulfated glycosaminoglycan was administered IM twice a week for a total of 8 treatments to all dogs with osteoarthritis and to clinically normal control dogs.

Results—Lameness scores after PSGAG treatment in osteoarthritic dogs improved in 12 of the 16 dogs. Serum COMP concentrations in osteoarthritic dogs were significantly higher than in control dogs before treatment. Lameness scores in osteoarthritic dogs decreased significantly after treatment, compared with before treatment. Lameness scores of 9 dogs with hind limb lameness improved significantly after treatment; these dogs had corresponding decreases in serum COMP concentrations. After treatment, serum COMP concentrations and lameness scores of 7 dogs with forelimb lameness remained high and were significantly higher than those of dogs with hind limb lameness. Serum MMP-9 activities of dogs with forelimb lameness were significantly higher than in dogs with hind limb lameness after treatment.


Osteoarthritis, also known as degenerative joint disease, is the end result of many articular disorders. It is a common arthropathy with radiographic features that include periosteal proliferation, deformity of the subchondral bone, and narrowing of joint spaces, mainly as a result of damage to articular cartilage. These changes are irreversible,1 are progressive, and result from joint instability or subluxation. The current treatment of osteoarthritis is aimed at slowing disease progression through alteration of pathobiological pathways.

Polysulfated glycosaminoglycan is a slow-acting disease-modifying agent that has been shown experimentally to slow and clinically ameliorate osteoarthritis.2,4 Polysulfated glycosaminoglycan is a semisynthetic polysulfated chondroitin sulfate (repeating sulfated residues of β-D-glucuronic acid and α-N-acetylgalactosamine linked by glycosidic β 1, 3 bonds) with approximately 3.25 sulfate groups/repeating disaccharide unit. The anionic charge and conformation of the drug are thought to enable its interaction with the active moiety in MMPs and thereby inhibit the activity of these enzymes. Results of in vitro and in vivo experiments have revealed the ability of supplemental PSGAG to diminish matrix molecule degradation3,5 and, in some studies, to enhance matrix molecule synthesis.6 The complete mechanism of the disease-modifying effects of PSGAG is not fully understood.

Cartilage oligomeric matrix protein is a macromolecule distributed abundantly in cartilage,7,8 synovi-
Cartilage oligomeric matrix protein is pentameric, consists of 5 globular domains that are attached to a central assembly domain by flexible strands and belongs to the thrombospondin family. Fragments of COMP have been detected in the diseased cartilage, synovial fluid, and serum of patients with osteoarthritis and rheumatoid arthritis. We have already reported that synovial fluid COMP concentration in dogs with naturally developing and experimentally induced arthropathy is higher than in dogs with normal joints and measurement of serum COMP concentration can be useful when differentiating among arthropathies in dogs. Matrix metalloproteinases are fundamental to the pathogenesis of joint disease and the breakdown of articular cartilage matrix, which occurs in diseases such as osteoarthritis and rheumatoid arthritis and is thought to occur primarily as a result of enzymatic degradation. Purified COMP is a substrate for MMPs including interstitial collagenase (MMP-1), collagenase-3 (MMP-13),stromelysin-1 (MMP-3), and gelatinase (MMP-2, and -9). Matrix metalloproteinase-2 and -9 have been examined for their possible role in joint disease in humans, dogs, and horses. Matrix metalloproteinase-2 and -9 exert a degradative action on collagens and proteoglycans in cartilage matrix and are able to degrade aggrecan in a similar manner to other MMPs, as well as cartilage link protein. Matrix metalloproteinase-2 and -9 concentrations are increased in synovial fluid from diseased joints. Matrix metalloproteinase-9 is produced by chondrocytes, peripheral blood monocytes, and neutrophils, and MMP-2 is produced by chondrocytes and synovial fibroblasts. Thus, MMP-2 and -9 are potential targets for therapeutic investigation in the treatment of joint disease. The purpose of the study reported here was to investigate the effects of treatment with PSGAG on serum COMP and CRP concentrations, serum MMP-2 and -9 activities, and lameness in dogs with arthropathy.

**Materials and Methods**

**Animals and sample collection**—Polysulfated glycosaminoglycan (5 mg/kg) was administered IM to 16 privately owned dogs that were brought to the Veterinary Teaching Hospital of Kagoshima University because of arthropathy (osteoarthrosis) and had chronic lameness and osteophytes on radiographic evaluation of the affected joint and also to 5 clinically normal control dogs in our laboratory. Informed consent was obtained from owners prior to the use of dogs in the study. Polysulfated glycosaminoglycan administration was performed twice a week for 4 weeks for a total of 8 treatments. Blood samples were obtained just before PSGAG administration and the day after the final administration. After centrifugation, serum samples were stored at −80°C until analysis. Also, all dogs were subjected to physical examination, observation of limb use, and radiographic examination. The study was approved by the Kagoshima University Animal Care and Use Committee.

**Lameness score**—All dogs were evaluated for lameness and weight bearing on the basis of criteria reported previously with some modifications. The lameness scale was as follows: 1 = standing with weight bearing and walking normally; 2 = standing with weight bearing normally and slight lameness while walking; 3 = standing with weight bearing normally and severe lameness while walking; 4 = incomplete weight bearing at standing and lameness while walking; and 5 = non-weight bearing at standing and also during walking.

**Preparation of canine COMP**—Canine COMP was purified by use of a method described elsewhere. Briefly, 10 g (wet weight) of frozen cartilage slices that was obtained from normal hip joints of an adult dog euthanized for reasons unrelated to orthopedic disease was milled in liquid nitrogen to obtain a fine particulate. Powdered tissue were suspended in 10 volumes of 4M guanidine hydrochloride in 50mM sodium acetate buffer (pH, 5.8) containing proteinase inhibitors (10mM EDTA, 100mM 6-aminohexanoic acid, 5mM benzamidine HCl, and 5mM N-ethylmaleimide), homogenized, and extracted at 4°C for 24 hours with gentle stirring. After centrifugation (17,000 X g at 4°C for 30 minutes), supernatant was collected, filtered, and dialyzed against a solution containing 7M urea, 20mM Tris HCl (pH, 8.0), 5mM EDTA, 1mM benzamidine, and 2mM N-ethylmaleimide. This solution was applied to an anion-exchange column. Material bound to the column was eluted with a linear gradient of 0 to 1.00M NaCl in the same buffer. A peak fraction eluted between 0.2 and 0.3M NaCl; therefore, those fractions were pooled to provide a COMP-rich fraction, which was dialyzed against distilled water. The preparation was considered to include cartilage COMP on the basis of analysis by use of SDS-PAGE and western immunoblot analysis that used a monoclonal antibody to human cartilage COMP (ie, 17C10). The preparation was subsequently lyophilized and used as a canine COMP standard.

Sodium dodecyl sulfate-polyacrylamide gel electrophoresis was performed in accordance with the Laemmli protocol. Nonreduced and reduced COMP molecules were applied to gradient (4% to 12%) gels. After electrophoresis, gels were stained with Coomassie brilliant blue R-250 or electrotransferred onto polyvinylidene difluoride membranes in Tris-glycine buffer (25mM Tris HCl, 192mM glycine [pH, 8.3], and 20% methanol) at 50 V overnight at 4°C. The transferred membrane was blocked and then analyzed by use of a murine monoclonal antibody (ie, 17C10) diluted 1:1,000 in 2% skim milk in Tris-buffered saline (0.9% NaCl) solution containing 0.05% Tween 20. Positive binding was detected by use of alkaline phosphatase–conjugated goat anti-mouse IgG antibody diluted 1:10,000 in 2% skim milk in Tris-buffered saline solution containing 0.05% Tween 20 and developed by use of 5-bromo-4-chloro-3-indolyl phosphate nitroblue tetrazolium as a substrate.

**Serum COMP analysis**—An inhibition ELISA for canine COMP was performed with the method described elsewhere. Fifty microliters of purified canine COMP antigen in a coating buffer (20mM sodium carbonate, 20mM sodium bicarbonate, and 0.05% sodium azide [pH, 10]) was placed into each well of 96-well ELISA plates, incubated for 2 hours at room
temperature (approx 25°C), and incubated overnight at 4°C. Canine COMP standards were prepared as serial 2-fold dilutions in PBS solution containing 0.05% Tween 20 (pH 7.0). Samples of serum were diluted in the same buffer. Aliquots (70 µL) of diluted standards (final dilutions, 40,000 to 156.3 ng/mL) and serum samples (final dilutions, 1:10 and 1:20) were mixed with the same volume of monoclonal antibody 17C10 (final dilution, 1:20,000 in PBS solution containing 0.05% Tween 20 [pH, 7.0]) and then incubated overnight at 4°C.

Coated wells were washed, and 100 µL of the inhibition mixture was added to each well. Plates were incubated for 1 hour at room temperature and then for 1 hour at 4°C. After incubation, plates were washed and 100 µL of alkaline phosphatase-conjugated goat antimouse IgG diluted 1:10,000 in PBS solution containing 0.05% Tween 20 was added to each well. Plates were incubated again and then washed, and 100 µL of glycine buffer (0.1M glycine, 1mM MgCl₂, and 0.5% Triton X-100 in 50mM Tris-HCl [pH, 7.5]). Gels were incubated for 18 hours at 37°C. After electrophoresis of a mixture of 5 µL of serum (diluted 3:100 with distilled water) and 5 µL of zymography buffer (4% sodium dodecyl sulfate, 20% glycerol, 1% bromophenol blue, and 2.5M Tris-HCl [pH, 6.8]), the sodium dodecyl sulfate was removed from the gels by several washes with 0.1% Triton X-100 in 50mM Tris-HCl (pH, 7.5). Gels were then rinsed with distilled water, and subsequently, the zymograms were developed for 18 hours at 37°C in re- folding buffer (50mM Tris-HCl [pH, 7.5], 0.15M NaCl, 10mM CaCl₂, and 0.02% NaN₃). Gels were stained with 1% Coomassie brilliant blue for 30 minutes and then destained in distilled water overnight. A mixture of 5 µL of canine MMP-2 and -9 standard (diluted 3:10 with distilled water) from a lipopolysaccharide-stimulated canine peripheral blood mononuclear cell culture and 5 µL of zymography buffer was loaded into 1 well/plate as the internal standard for serum MMP-2 and -9 activities.

Enzyme activities of samples were quantified as the net intensity of the gelatinolytic activity including the bands of pro- and active MMP-2 and -9 in each lane, which were determined on the basis of the molecular size of those bands (canine pro-MMP-2, 72 kd; active-MMP-2, 67 kd; pro-MMP-9, 92 kd; and active MMP-9, 83 kd) by use of software packages. Actual measurements for samples were converted to values relative to the MMP-2 and -9 standards. To determine the intra- and interassay variability, fresh aliquots of serum were thawed and 10 measurements were repeated on a single plate (1 gel) for 10 consecutive working days (ie, total of 10 gels).

**Results**

**Lameness score**—Lameness score after PSGAG treatment in 12 of 16 osteoarthritic dogs improved significantly (P < 0.001; Table 1). In 4 dogs with osteoarthritis, scores did not improve but signs of limb pain did not worsen. Lameness scores in osteoarthritic dogs before and after treatment were 3 (3 to 5) and 1 (1 to 4), respectively. Lameness scores in 9 dogs with hind limb lameness improved from 3 (2 to 5) to 1 (score of 1 for all dogs) with treatment, whereas lameness scores in 7 dogs with forelimb lameness did not change (from 3 [3 to 4] to 3 [1 to 4]). A significant (P < 0.001) difference was found in posttreatment lameness scores between hind limbs and forelimbs (Figure 1).
Serum COMP concentration—Serum COMP concentrations in osteoarthritic dogs were significantly (P = 0.019) higher (18.32 [interquartile range, 7.50 to 24.23] µg/mL) than in control dogs (12.06 [10.16 to 14.54] µg/mL) before treatment and decreased significantly (P < 0.001) after PSGAG treatment in osteoarthritic dogs (13.87 [6.09 to 22.48] µg/mL), compared with before treatment (Table 1). Serum COMP concentrations in 7 dogs with forelimb lameness before (20.7 [15.20 to 24.23] µg/mL) and after (16.65 [11.62 to 22.48] µg/mL) treatment were significantly higher than values in 9 dogs with hind limb lameness before (15.05 [7.50 to 23.84] µg/mL; P = 0.009) and after (12.32 [6.09 to 17.36] µg/mL; P = 0.016) treatment (Figure 2). Also, in the 12 osteoarthritic dogs that had an improvement in lameness score, serum COMP concentrations before and after treatment were 16.29 (7.50 to 24.23) µg/mL and 12.52 (6.09 to 17.36) µg/mL, respectively, which represented a significant (P = 0.028) decrease in response to treatment. In the 4 osteoarthritic dogs that had no improvement in lameness score, serum COMP concentrations before and after treatment were 20.98 [15.05 to 24.23] µg/mL and 22.48 [15.20 to 24.23] µg/mL, respectively.
Polysulfated glycosaminoglycan is a slow-acting disease-modifying agent that has been shown experimentally to slow the development of degenerative joint disease and is used in veterinary practice for treating horses and dogs with joint disease. Unlike steroid and nonsteroidal anti-inflammatory drugs, PSGAG does not exert an instantaneous effect but is used widely because of its high level of safety and the possibility of long-term administration with few adverse effects. The mechanism of action of PSGAG is considered to be an acceleration of extracellular matrix production by stimulation of chondrocytes and synoviocytes, inhibition of proteolytic enzymes, and the production offlammatory mediators. Polysulfated glycosaminoglycan has potential use for pain modification in arthropathy and for inhibition of cartilage matrix degradation. The pharmacologic action of PSGAG appears to involve the repair of damaged joint cartilage, inhibition of proteoglycan degradation, and an anti-inflammatory effect when distributed to damaged articular cartilage, protecting against loss of proteoglycan from the articular cartilage. Also, PSGAG inhibits the degradation of articular cartilage and inflammation of periarticular tissue through inhibition of MMP activity, collagenolytic enzyme activity, or both as well as inhibition of nitric oxide production from inflammatory cells.

Administration of PSGAG is a useful adjunctive treatment for osteoarthritis in dogs and horses but is not a panacea. Evidence exists that PSGAG is beneficial for the prevention and treatment of experimentally induced osteoarthritis and modifying the progression of degenerative changes in canine hip dysplasia. In our study, an improvement in lameness scores was detected in 12 of 16 osteoarthritic dogs. Improvements in lameness became evident from the fourth administration of PSGAG onward, and no adverse effects were observed. Cartilage oligomeric matrix protein is a prominent noncollagenous component of cartilage, contributing to approximately 1% of the wet weight of articular tissue. Collagenous structure breakdown in cartilage tissue accompanies degradation of COMP and increases of COMP fragment concentrations in synovial fluid. Fragments of COMP have been detected in diseased cartilage, synovial fluid, and serum of patients with primary osteoarthritis, synovitis, and rheumatoid arthritis. Measurements of synovial fluid and serum COMP concentrations in dogs indicate that COMP concentrations can be used to diagnose cartilage degradation in arthropathy; serum COMP concentrations increase after increases in synovial fluid COMP concentrations in experimentally induced synovitis. In our study, the pretreatment serum COMP concentrations in dogs with osteoarthritis were significantly higher than in control dogs. Serum COMP concentrations in osteoarthritic dogs decreased significantly after PSGAG treatment, along with an improvement in lameness. Also, serum COMP concentrations were significantly higher in dogs with forelimb lameness than in dogs with hind limb lameness, and the response to PSGAG treatment for forelimb lameness was less pronounced than for hind limb lameness. Our results indicate that destruction of collagen in cartilage in osteoarthritic dogs was inhibited by PSGAG administration and that the potential effect of PSGAG would be more evident in hind limb lameness.

Matrix metalloproteinase-2 and -9 are collagenolytic enzymes; MMP-2 is formed constitutively with collagen turnover. Matrix metalloproteinase-9 processes fragmentated matrix components that are induced in response to tissue injury and inflammation. Results of previous studies indicate that MMP-2 and -9 activities in synovial fluid are high in joints with osteoarthritis and rheumatoid arthritis and can be inhibited by PSGAG. However, for our results, only serum MMP-9 activity in dogs with forelimb lameness was significantly higher than in dogs with hind limb lameness after treatment. The finding that serum MMP-9 activity did not decrease after PSGAG treatment indicates that concentrations or activities of other inflammatory mediators (eg, transforming growth factor-β1) continued to be high in affected joints, despite PSGAG treatment. Serum COMP concentration is significantly associated with joint inflammation and synovitis. In our study, although the difference was not significant, the value of serum CRP concentration in dogs with forelimb lameness was higher than in dogs with hind limb lameness before and after treatment. In terms of the proportion of body weight borne by forelimbs and hind limbs in dogs, forelimbs carry a greater load. Therefore, forelimbs might be more susceptible to mechanical stress and joint inflammation and thus more difficult to improve with PSGAG treatment.
Results of our study indicate that PSGAG might be effective for improving cartilage COMP turnover and joint inflammation and that the observed changes reflect serum COMP concentrations. Lameness scores were improved in parallel with changes in serum COMP concentrations. We conclude that IM administration of PSGAG is effective for inhibition of COMP degradation with improvement in lameness and that one of the mechanisms of action of PSGAG is inhibition of COMP degradation.

References