

Use of tissue Doppler imaging to confirm the diagnosis of dilated cardiomyopathy in a dog with equivocal echocardiographic findings

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- ▶ At present, diagnosis of the preclinical phase of dilated cardiomyopathy in dogs remains a challenge for veterinarians.
- ▶ Tissue Doppler imaging is a recently developed ultrasonographic technique that allows quantification of regional myocardial function in real time by measurement of myocardial velocities.
- ▶ In dogs with equivocal conventional echocardiographic findings of dilated cardiomyopathy, severe myocardial alterations may be detected via tissue Doppler imaging.

A 1-year-old 42-kg (92.4-lb) sexually intact female Great Dane was referred to the Cardiology Unit of Alfort for echocardiographic examination prior to anesthesia for surgical correction of prolapse of the right third eyelid gland. The dog had no history of clinical signs related to disease of the respiratory or cardiovascular system. Findings of a physical examination were within normal limits.

By use of an ultrasound machine^a equipped with a 2.5- to 3.5-MHz phased-array transducer, 2-dimensional (2D) and M-mode echocardiography, color-flow imaging, spectral Doppler examination, and tissue Doppler imaging (TDI) were performed by the same trained observer (VC) with continuous ECG monitoring. For all ultrasonographic examinations, the dog was awake and gently restrained in a standing position; this method has already been proven by our group to have good repeatability and reproducibility for conventional echocardiography¹ as well as TDI examination.^{2,b,c}

For 2D color TDI examination, real-time color Doppler was superimposed on the gray scale with a frame rate ≥ 100 frames/s. The Doppler gain was adjusted to maintain optimal coloring of the myocardium, and Doppler velocity range was set as low as possible to avoid aliasing. Left ventricular free wall (LVFW) velocities resulting from the radial left ventricular motion were measured in the right parasternal ventricular short-axis view between the 2 papillary muscles, as previously described.^{2,c} The angle of the beam was carefully aligned to be perpendicular to the LVFW. Measurements were made on an endocardial

and an epicardial segment (2×2 mm) of the LVFW. Simultaneous endocardial and epicardial velocity profiles were obtained by use of a stand-alone, off-line measuring system.^d

During this initial evaluation of the dog (day 0), conventional M-mode and 2D echocardiographic parameters^{3,5} were assessed (Table 1). No left ventricular or left atrial dilatation was observed on the initial echocardiogram. The left ventricular and septal systolic thickenings were normal in appearance. The ratio of the pre-ejection period (40 milliseconds) to left ventricle ejection time (160 milliseconds) was not greater than that expected for a clinically normal dog. The E point-to-septal separation distance and the shortening fraction were at the upper and lower reference limits, respectively. The index of sphericity (ratio of the left ventricle diastolic length to the M-mode left ventricle diastolic dimension⁶) was also considered equivocal, although the sensitivity and specificity of this parameter are still unknown; in the dog of this report, the index of sphericity was 1.43, and index values < 1.65 are considered to represent increased sphericity. Spectral Doppler imaging of the mitral inflow confirmed that the ratio of the diastolic E wave to the diastolic A wave (E:A ratio) was within the normal range (E:A ratio, 1.5; reference range, 1.04 to 2.42⁷), and that the E wave deceleration time was not decreased (90 milliseconds; normal lower limit, 80 milliseconds⁶), compared with that expected in clinically normal dogs. Very rarely, isolated unifocal left ventricular premature beats (0 to 2 beats/min) with a normal heart rate (112 beats/min; reference range, 100 to 130 beats/min³) were observed on ECG monitoring. At this point of the evaluation on day 0, the shortening fraction, sphericity index, E point-to-septal separation distance, and presence of ventricular premature beats were all considered to be evidence of possible dilated cardiomyopathy (DCM). However, according to the proposed ultrasound criteria for the diagnosis of canine DCM,⁶ the total score for this dog was 4 (3 points for the slight decrease in the index of sphericity and 1 for the rare ventricular premature beats), whereas a total minimum score of 6 is needed to confirm DCM.

During the evaluation of the dog on day 0, a TDI examination was also performed. As previously described,^{2,c} the radial velocity profiles included 1 positive systolic wave and 2 negative diastolic waves (E and A, in early diastole and late diastole, respectively; Figure 1). However, compared with reference values, systolic and early diastolic endocardial velocities were lower than normal, leading to a marked decrease in the 2 corresponding

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Table 1—Conventional M-mode and 2-dimensional echocardiographic parameters measured in a Great Dane during the initial evaluation (day 0) and at reevaluation 4 months later.

Echocardiographic parameters	Day 0 (weight, 42 kg [92.4 lb])	Normal ranges for day-0 values*	Reevaluation (weight, 47 kg [103.4 lb])	Normal ranges for reevaluation values*
Left ventricular end-diastolic diameter (mm)	43.5	46.2–51.2 ⁴	62.3	48.7–54.4 ⁴
Left ventricular end-systolic diameter (mm)	32.0	29.0–32.6 ⁴	46.8	30.7–34.8 ⁴
Interventricular septal systolic thickening (%)	38.5	34–73.2 ²	35.6	34–73.2 ²
LVFW systolic thickening (%)	56.1	45.7–75.9 ⁵	40.8	45.7–75.9 ⁵
Shortening fraction (%)	28.5	18–36 ³	24.6	18–36 ³
Left atrium size (mm)	22.8	27–30 ⁴	41.3	28.4–32.0 ⁴
Aorta diameter (mm)	27.8	28.8–31.2 ⁴	32.1	30.6–33.4 ⁴
Left atrium size-to-aorta diameter ratio	0.82	0.83–1.13 ³	1.28	0.83–1.13 ³
E point-to-septal separation (mm)	10.6	5.0–12.0 ²	9.9	5.0–12.0 ²
Pre-ejection period-to-left ventricular ejection time ratio	0.20	0.10–0.54 ⁵	0.32	0.10–0.54 ⁵

*Normal ranges for Great Danes that weigh 52 to 75 kg (114.4 to 165.0 lb) have been described.² However, no specific reference ranges are available for young Great Danes < 52 kg. Therefore, the normal ranges of left ventricular and atrial diameters, aortic diameters, and myocardial wall thicknesses indicated are those described in adult dogs of 42 kg (day 0) and 47 kg (at reevaluation).⁴ Reference ranges for derived echocardiographic parameters are those reported by Koch et al³ or by Boon et al⁵ if data were not available for Great Danes.

LVFW = Left ventricular free wall.

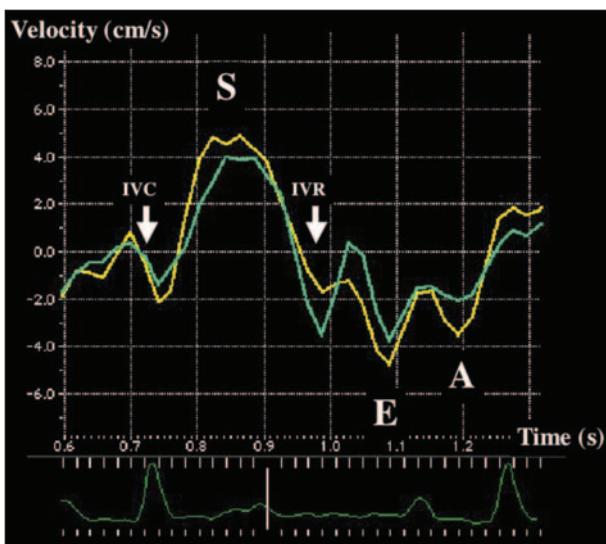


Figure 1—Analysis of left ventricular free wall (LVFW) radial motion during the initial echocardiographic evaluation (day 0) of a Great Dane by use of 2-dimensional color tissue Doppler imaging in the right parasternal short-axis view. The yellow and green curves (endocardial and epicardial velocity profiles, respectively) are nearly superimposed during the whole cardiac cycle, indicating the lack of systolic and diastolic myocardial velocity gradients. The trace at the bottom of the image represents the corresponding ECG. IVC = Isovolumic contraction phase. S = Peak velocity of the LVFW during systole. IVR = Isovolumic relaxation phase. E = Peak velocity of the LVFW during early diastole. A = Peak velocity of the LVFW during late diastole.

myocardial velocity gradients (Table 2). Moreover, the E:A ratio was decreased particularly in the endocardial segment. Late diastolic endocardial and epicardial velocities were not decreased, and the corresponding velocity gradient was within the normal range. On the basis of these TDI data, it was concluded that the dog had a severe systolic and early diastolic myocardial alteration, which was highly suggestive of DCM.⁶ Because of this, the dog did not undergo surgical repair of the prolapsed third eyelid and the owner was advised to return to the dog within the next 4 to 6 months for reevaluation of both conventional and TDI echocardiographic parameters.

Four months later, the dog was reevaluated as planned. The owner noticed progressive exercise intolerance since the last examination. On physical examina-

Table 2—Radial tissue Doppler imaging (TDI) parameters measured in the endocardial and epicardial layers of the LVFW of a Great Dane at initial evaluation (day 0).

TDI parameters (cm/s)	Day 0	Reference ranges (cm/s) ⁸
S wave (endocardial)	4.4	6.0 ± 1.01
S wave (epicardial)	3.9	3.4 ± 0.77
Systolic velocity gradient (S _{endocardial} - S _{epicardial})	0.5	2.5 ± 0.97
E wave (endocardial)	5.4	8.1 ± 1.82
E wave (epicardial)	5.3	3.5 ± 0.78
Early diastolic velocity gradient (E _{endocardial} - E _{epicardial})	0.1	4.5 ± 1.78
A wave (endocardial)	5.3	3.7 ± 0.99
A wave (epicardial)	2.8	1.7 ± 0.65
Late diastolic velocity gradient (A _{endocardial} - A _{epicardial})	2.5	1.9 ± 0.70

S wave = Peak velocity of the LVFW during systole. E wave = Peak velocity of the LVFW during early diastole. A wave = Peak velocity of the LVFW during late diastole.

tion, capillary filling time and the color of the mucous membranes were considered normal. However, the dog was tachypneic (≥ 50 breaths/min). Auscultation revealed the presence of paroxysmal tachycardia that was associated with a weak arterial pulse. Thoracic radiography revealed left-sided cardiomegaly without pulmonary edema. Several episodes of unifocal ventricular tachycardia (heart rate > 220 beats/min) were observed on an ECG and also during the ultrasonographic examination. While echocardiographic measurements were made, the evaluator (VC) ensured that ventricular tachycardia was not present (Table 1). Conventional echocardiography confirmed a marked systolic and diastolic left ventricular dilatation associated with a left atrial dilatation (41.3 vs 22.8 mm at day 0; reference range, 28.4 to 32.0 mm⁴; Figure 2). The index of sphericity had decreased dramatically, indicating a marked rounding of the left ventricle (0.98 vs 1.42 at day 0). By use of the proposed criteria for diagnosis of DCM in dogs,⁶ the total score for the dog was 8 (3 points for the decrease in the index of sphericity, 3 for the left ventricular dilatation, 1 for the ventricular arrhythmias, and 1 for the left atrial dilatation); thus, the diagnosis of DCM was confirmed.

Results of previous studies in humans and various animal models of human heart diseases have indicated that TDI is a sensitive technique for the diagnosis of subtle regional myocardial dysfunction, such as

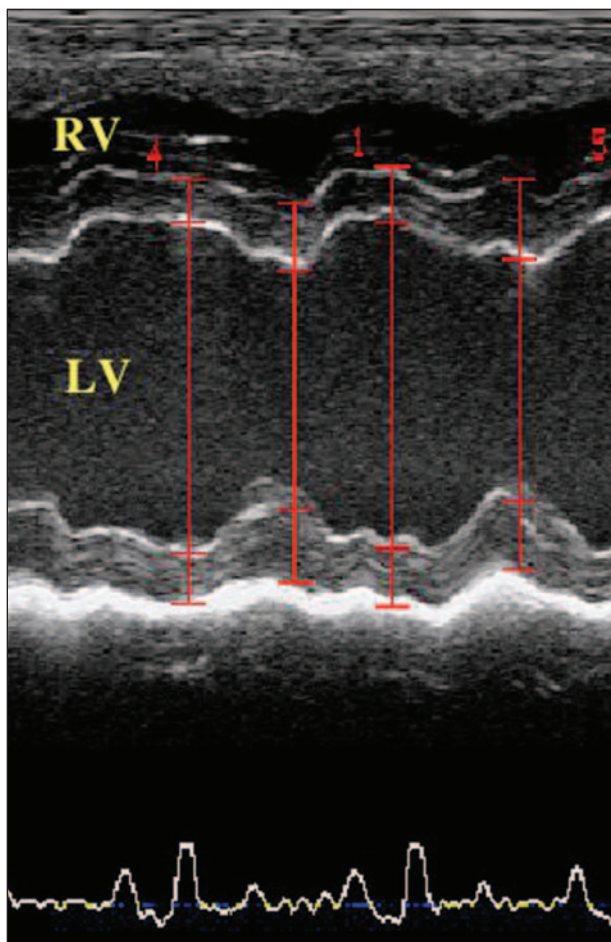


Figure 2—Left ventricular M-mode echocardiographic image obtained from the dog in Figure 1 four months after the day-0 evaluation. Notice the systolic and diastolic left ventricular dilatation. The trace at the bottom of the image represents the corresponding ECG. LV = Left ventricle. RV = Right ventricle.

ischemia,^{8,9} heart transplant disorders,¹⁰ and cardiomyopathies. The myocardial disorders that have been studied most via TDI are primary or secondary **hypertrophic cardiomyopathies (HCM)** in human patients^{11,12} and also in animal models of HCM involving rats and rabbits.^{13,14} In a study involving a transgenic rabbit model of human HCM, Nagueh et al¹⁴ reported that TDI accurately identified the mutant transgenic rabbits irrespective of the extent of cardiac hypertrophy; via TDI, all mutant rabbits (even those without left ventricular hypertrophy) had significantly reduced systolic and diastolic myocardial velocities, which indicated that TDI was more sensitive for detection of moderate myocardial alterations than were conventional echocardiographic methods. Similar findings have been reported for humans with familial HCM; myocardial contraction and relaxation velocities detected via TDI were reduced in patients with familial HCM, including those without left ventricular hypertrophy.¹² The detection of reduced myocardial contraction and relaxation velocities via TDI had a sensitivity of 100% and a specificity of 93% for identifying human patients with mutations in the absence of left ventricular hypertrophy.¹²

At present, conventional echocardiography is used to screen for DCM in humans and animals, and the diagnosis is made on the basis of detection of reduced inotropism, dilatation of cardiac chambers, and increased sphericity.⁶ However, these ultrasonographically detectable functional and morphologic changes are sometimes absent depending on the stage of the disease,⁶ and diagnosis of the preclinical phase of DCM in dogs (occult DCM¹⁵) still remains a challenge. In another study⁶ by our group, the accuracy of TDI for detection of early myocardial dysfunction associated with DCM in Golden Retrievers with muscular dystrophy was demonstrated. Because of a decrease in the endocardial-epicardial gradient,¹⁶ the dogs with the dystrophin mutation were detected via TDI before development of congestive heart failure and myocardial dysfunction, as determined by conventional echocardiographic parameters (ie, left ventricular dimensions and shortening).⁶ However, to our knowledge, the accuracy of TDI for early detection of naturally occurring DCM in dogs in a clinical setting has not been reported. In the dog of this report, the systolic and diastolic radial myocardial motion was markedly altered 4 months before development of unequivocal signs of DCM. This finding, which needs to be confirmed in other dog breeds and in a larger number of animals, provides new insight into the pathogenesis of DCM; it also suggests that TDI may be a sensitive technique of screening for DCM during the preclinical phase in dogs, which is not detectable via currently used conventional echocardiographic techniques. At present, dobutamine stress echocardiography is a method of determining the presence of preclinical systolic and diastolic myocardial dysfunction.¹⁷ This technique has been evaluated in healthy Doberman Pinschers to determine which parameters were associated with the development of occult DCM.¹⁸ Further studies are required to compare the sensitivities of TDI and dobutamine stress echocardiography for identification of occult cardiomyopathies in dogs. The preclinical diagnosis of cardiomyopathies in dogs could enable veterinary practitioners to institute medical treatment early and thereby prevent the development of heart failure; it could also help to identify dogs carrying a mutation for DCM, enabling exclusion of those individuals from breeding programs.

^aVingmed system 5, General Electric Medical System, Waukesha, Wis.

^bChetboul V, Athanassiadis N, Carlos C, et al. Quantification of longitudinal left ventricular motion using tissue Doppler imaging: new indices of myocardial function in dogs (abstr). *J Vet Intern Med* 2003;17:399.

^cChetboul V, Athanassiadis N, Carlos C, et al. Quantification of radial left ventricular motion in healthy dogs using tissue Doppler imaging: intraday and interday variability (abstr). *J Vet Intern Med* 2003;17:440.

^dEcho Pac 5.4 software for system 5, GE-Vingmed Ultrasound, Waukesha, Wis.

^eChetboul V, Escricou C, Blot S, et al. Early detection of regional myocardial function alterations in a dog model of dilated cardiomyopathy by tissue Doppler imaging study (abstr). *Circulation* 2001;104(suppl 17):351.

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Selected abstract for JAVMA readers from the American Journal of Veterinary Research

Serum concentrations of cortisol, sex hormones of adrenal origin, and adrenocortical steroid intermediates in healthy dogs following stimulation with two doses of cosyntropin

Linda A. Frank et al

Objective—To compare the effects of 2 doses of cosyntropin (5 µg/kg vs 250 µg, IV) on serum concentrations of cortisol, sex hormones of adrenal origin, and adrenocortical steroid intermediates and determine the optimal sample collection time after adrenal stimulation with cosyntropin.

Animals—10 healthy, privately owned, neutered dogs.

Procedure—Dogs were randomly assigned to initially receive cosyntropin at 5 µg/kg or as a total dose of 250 µg, IV. Dogs received the alternate dose 1 to 2 weeks later. Serum was obtained from blood samples collected before (0 minutes) and 30, 60, 90, and 120 minutes after cosyntropin administration.

Results—Maximum stimulation of cortisol, androstenedione, progesterone, and 17-hydroxyprogesterone production was achieved at 60 minutes following IV administration of cosyntropin at 5 µg/kg or as a total dose of 250 µg. Serum estradiol concentration did not increase in response to either cosyntropin dose. For all hormones, no significant difference in serum hormone concentrations was found among sample collection times of 0, 30, 60, and 90 minutes when comparing the 2 doses of cosyntropin.

Conclusions and Clinical Relevance—Cosyntropin, when administered at 5 µg/kg, IV, effectively stimulated maximum production of cortisol, sex hormones of adrenal origin, and adrenocortical steroid intermediates at 1 hour after administration. (*Am J Vet Res* 2004;65:1631–1633)



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