Computed tomography lymphangiography via intrametatarsal pad injection is feasible in cats with chylothorax

Chuan Chiang, DVM; Kuan-Sheng Chen, PhD, MANZCVS, DVM; Hsien-Chieh Chiu, MS, DVM; Cheng-Shu Chung, PhD, DVM; Lee-Shuan Lin, PhD, DVM*

1UniCore Animal Hospital, Taipei City, Taiwan
2Department of Veterinary Medicine, College of Veterinary Medicine, National Chung Hsing University, Taichung, Taiwan
3Veterinary Medical Teaching Hospital, College of Veterinary Medicine, National Chung Hsing University, Taichung, Taiwan
4Tzuoo Ann Animal Hospital, New Taipei City, Taiwan
5Laboratory of Veterinary Surgery, Department of Veterinary Medicine, College of Veterinary Medicine, National Pingtung University of Science and Technology, Pingtung, Taiwan
6Laboratory of Veterinary Diagnostic Imaging, Department of Veterinary Medicine, College of Veterinary Medicine, National Pingtung University of Science and Technology, Pingtung, Taiwan
7Veterinary Medical Teaching Hospital, College of Veterinary Medicine, National Pingtung University of Science and Technology, Pingtung, Taiwan

*Corresponding author: Dr. Lin (linleeshuan@gmail.com)

https://doi.org/10.2460/ajvr.21.10.0163

OBJECTIVE
To evaluate the feasibility of CT lymphangiography via intrametatarsal pad injection in cats with chylothorax.

ANIMALS
7 client-owned cats.

PROCEDURES
This was a multicenter, retrospective, descriptive study. Medical records and imaging data from 4 veterinary hospitals were reviewed to identify cats with chylothorax that had undergone intrametatarsal pad injection via CT lymphangiography. In total, 7 client-owned cats were included in the study. Signalment, history, image findings, and follow-up data were recorded. Descriptive statistics were used to analyze the success rate of thoracic duct (TD) enhancement and describe relevant clinical findings.

RESULTS
Enhancement of TDs was successful in 6 of the 7 cats within 5 to 15 minutes after initiating intrametatarsal pad injection under general anesthesia. Successful migration of contrast medium into the lymphatic vessels cranial to the popliteal lymph nodes was observed in all cats within 5 minutes after injection. The recommended dose of contrast medium to achieve TD enhancement was 1 mL/kg (0.5 mL/kg/pad; concentration, 350 mg of iodine/kg). Only 1 cat had mild swelling of the paws after the procedure, and it recovered quickly without pain medication; no cats experienced lameness. Similar to dogs and unlike in previously published reports, 72% of TD branches were located in the right hemithorax.

CLINICAL RELEVANCE
CT lymphangiography via intrametatarsal pad injection is a feasible and safe procedure for cats with chylothorax. This technique provides detailed information regarding the unique TD anatomy and cisterna chyli location. It also contributes to surgical planning.

In cats, chylothorax is caused by an abnormal flow or pressure in the thoracic duct (TD) and its branches, leading to chylous accumulation in the thoracic cavity.1 Multiple possible etiologies, including cardiomyopathy, neoplasia, congenital cardiac abnormalities, thrombosis, lung lobe torsion, and dirofilariasis, have been determined in feline chylothorax, but most cases are considered idiopathic.2–9 Surgical intervention is necessary if medical management is ineffective or impractical.1 Surgical intervention is warranted in animals with idiopathic chylothorax or those that are not responsive to medical management.1 Among surgical options, TD ligation, subtotal pericardiectomy, cisterna chyli ablation, thoracic embolization, and thoracic omentalization are commonly considered effective.5,10–13 In cats, a combination of TD ligation with subtotal pericardiectomy has a higher success rate (73%) for clinical resolution than performing TD ligation alone (41%).14,15 However, unlike the improved success rate when cisterna chyli ablation is combined with TD ligation in dogs (83%), 1 study14 indicated that the strategy did not result in better outcomes in cats.
Owing to the high variation of lymphatic vessels among individuals, identifying their position and branches during TD ligation is important. Intraoperative lymphangiography is commonly performed by injection of methylene blue or indocyanine green via mesenteric lymph node injection during laparotomy or feeding cream or corn oil before induction. Preoperative CT lymphangiography is helpful for surgical planning and determining the approach site. Ultrasound-guided injection of contrast medium into the mesenteric or popliteal lymph nodes in chylothorax cases has been reported. Nevertheless, these techniques have failed in some feline cases because of the difficulty of injecting large amounts of contrast medium into the small lymph nodes of cats.

Simpler alternative methods for CT lymphangiography by SC injection of the contrast medium into perianal tissue or the dorsal metatarsal region have been implemented in dogs. However, it may be difficult to inject contrast medium into the small perianal region of cats, and similar to dogs, infection is also a concern. Moreover, the potential needle trauma to the dorsal tendons and adjacent nerves during the injection because of the thin subcutis at the dorsal metatarsal region, particularly in small-sized animals, cannot be ignored. We previously reported the feasibility of CT lymphangiography via intrametatarsal pad injection as a safe and straightforward method for dogs. In this method, the thick subcutaneous adipose tissue of the pad allows for the injection of large amounts of contrast medium, enabling satisfactory visualization of lymphatic vessels, cisterna chyli, and the TD. This retrospective study aimed to assess the outcomes of CT lymphangiography via intrametatarsal pad injection in cats with chylothorax and to describe related clinical findings.

**Materials and Methods**

**Case selection**

This was a multicenter, retrospective, descriptive study. We reviewed medical records and imaging data obtained between July 1, 2018, and November 30, 2020, from the Veterinary Medical Teaching Hospital of National Pingtung University of Science and Technology (Pingtung, Taiwan), National Chung Hsing University Veterinary Medical Teaching Hospital (Taichung City, Taiwan), Tzuoo Ann Animal Hospital (New Taipei City, Taiwan), and UniCore Animal Hospital (Taipei City, Taiwan). Each hospital approved data usage. The inclusion criteria were client-owned cats with chylothorax that underwent CT lymphangiography via intrametatarsal pad injection with nonionic iodinated contrast medium. Patient data were excluded if they previously had TD surgery. This work involved the use of nonexperimental animals alone.

**Imaging equipment**

Four helical multidetector CT scanners with different imaging techniques were used. Two 16-slice multidetector CTs and two 64-slice multidetector CTs were used. The CT parameter details are listed elsewhere (Supplementary Appendix S1).

**CT scan and intrametatarsal pad lymphangiography**

Animals were anesthetized and maintained on gas anesthesia and subsequently positioned in sternal recumbency for CT scan. After location acquisition, helical precontrast scanning was performed. Lymphangiography was performed after aseptic preparation of the pads. The cats were injected with a nonionic iodinated contrast medium (iohexol [Omnipaque 350], 350 mg of iodine/kg; or ioversol [Optiray 350], 350 mg of iodine/kg) into the right and left hind limb metatarsal pads in equal aliquots with 21- or 22-gauge needles for 15 to 30 seconds. The metatarsal pads, distal hind limbs, and tarsal and thigh regions were massaged thoroughly from the distal to proximal direction for 3 to 4 minutes after the injection (Figure 1; Supplementary Video V1). The CT scan was performed 5 minutes after the intrametatarsal pad injection. If the contrast medium did not opacify the TD within 5 minutes of lymphangiography, the scans were repeated at 2- to 3-minute intervals until satisfactory TD enhancement was obtained. Postcontrast IV contrast CT scan was performed after lymphangiography with a dose of 2 mL/kg.

**Data recording**

**Clinical data**—Signalment (sex, breed, and body weight), CT imaging parameters, lymphangiography techniques used (contrast medium, injected volume, injection method, and intervals of delayed scans), adverse effects, and follow-up information were summarized from the medical records for each cat by 1 veterinarian (CC).

**Imaging analysis**—The CT images were reviewed in the soft tissue window (window level, 40 HU; window width, 400 HU) with commercial software (Osirix MD version 11.0.3; Pixmeo SARL). Qualitative and quantitative data were evaluated by 2 authors (CC and LSL). Observers could see the patient identification number and name but were unaware of the signalment, injected contrast volume, injection method, and outcome at the time of evaluation. The 3-D images were processed using a commercial imaging workstation (Attractive; PixSpace Ltd).

**Identification of lymphatic vessels and evaluation of TD enhancement**

Successful lymphangiography was defined as lymphatic vessel enhancement cranial to the popliteal lymph nodes. Enhancement of TD was deemed successful when the contrast medium enabled its identification. The number of visible TD branches was evaluated and recorded at each midvertebral region from T9 to L1. At the common TD ligation site at the T10 to T12 level, location of the TD branch relative to the aorta was expressed using a clockface analogy. The quality of TD enhancement at the T10 to T12 level was graded as follows: grade 3.
(excellent), grade 2 (good), grade 1 (fair), and grade 0 (poor). The distance between the TD and aorta was assessed. A line was drawn from the center of the aorta to the TD branches at the midvertebral region from T10 to T12, and the distance from the aortic wall to the TD branches was measured.

**Evaluation of the cisterna chyli**

The location of the cisterna chyli relative to the aorta and the pattern of the structure were assessed and recorded. The length of the cisterna chyli was also measured on sagittal planes.

**Evaluation of the efferent lymphatic pathway**

If the hind limbs and pelvic area were included in the scan, the pattern of the lymphatic efferent pathway was evaluated.

**Data analysis**

Statistical analyses were performed with a spreadsheet (Excel for Mac version 16.45; Microsoft Corp) and commercial software (Prism version 8.0; GraphPad Software Inc). A preliminary Shapiro-Wilk test was performed, and it demonstrated non-normally distributed data. Age and body weight were expressed as median and range values.

**Results**

**Study population**

Seven cats met the inclusion criteria and included 2 Ragdoll and 5 mixed-breed cats. The median age was 7 years (range, 2 to 15 years) and median body weight was 4.2 kg (range, 3.2 to 6.8 kg); 4 cats were spayed females, 2 were neutered males, and 1 was a sexually intact male.

**CT scan and intrametatarsal pad lymphangiography**

Prior to general anesthesia, thoracocentesis was performed in 5 cats, and the volume of aspirated chylous effusion ranged from 100 to 150 mL. The CT scanning included the thoracic, abdominal, and pelvic limb regions in 6 cats; 1 cat was only scanned from the thoracic inlet to the L4 level. One cat showed no TD enhancement even after repeated scans of lymphangiography. Therefore, 30 minutes after intrametatarsal pad injection, the scanning was discontinued.

The median dose of contrast medium for lymphangiography was 1.25 mL/kg (range, 1.0 to 2.0 mL/kg). The doses used in the 6 cases of successful TD enhancement were as follows: 1 mL/kg in 3 cats, 1.5 mL/kg in 1 cat, and 2 mL/kg in 2 cats. One cat was administered contrast medium at a dose of 1.5 mL/kg, but no TD enhancement occurred.

**Timing of lymphatic vessel and TD enhancement**

For all cats, lymphatic enhancement could be visualized cranial to the popliteal lymph node within 5 minutes after injection, indicating a 100% success rate of lymphangiography via intrametatarsal pad injection.

Enhancement of TD was identified in 6 of 7 cats within 15 minutes after injection. The TD was successfully identified within 5 minutes in 1 cat (14%) in the first scan and within 6 to 15 minutes by repeated scanning in 5 cats. No TD enhancement was found in 1 cat, in which the contrast medium accumulated in bilateral enlarged medial iliac lymph nodes.

**Quality and quantity of TD enhancement**

The quality of the TD enhancement at the T10 to T12 level was as follows: 4 cats exhibited grade 3 enhancement, 2 exhibited grade 2 enhancement, and 1 exhibited grade 0 enhancement. A fused 3-D image from the lymphangiography and angiography was created (Figure 2).

Multiple TD branches were identified in 6 cats with successful TD enhancement. The maximum numbers of TD branches were as follows: 2 in 3 cats, and 3 in the other 3 cats. The number of TD branches at different vertebral levels was variable (Supplementary Table S1).

A clockface analogy of the TD branches from a craniocaudal view at...
The distance between the TD and the aorta was measured at the T10 to T12 level. In almost all cats with successful TD enhancement (5/6 cats), the TDs were located adjacent to the aorta or less than 1 mm from it, except for 1 cat with a distance of 2.6 to 3.4 mm from the aorta at the T10 to T12 level.

Cisterna chyli location and structure

Successful cisterna chyli enhancement was noted in 6 cats, and the location ranged from L2 to L3. Five different types of cisterna chyli structures were identified, as follows: right to the aorta (n = 1 cat); dorsal (1); right and dorsal (1); right, dorsal, and ventral (2); and spiral (1; Figure 4). The median length of the cisterna chyli was 1.25 cm with a range of 1.0 to 3.3 cm.

Lymphatic pathway assessment

The hind limb and pelvic regions were included in the scanning field in 6 cats. This allowed for evaluation of the direct lymphatic efferent pathway from the popliteal lymph nodes to the medial iliac lymph nodes and the dorsal lymph vessel pathway coursing to the iliac lymph nodes along the gluteal region. Bilateral direct efferent pathways were observed in all 6 cats. Four cats had dorsal lymph vessel pathways as follows: 1 exhibited a bilateral dorsal lymph vessel pathway, whereas 3 exhibited right dorsal lymph vessel pathways (Figure 5).

Follow-up

Changes in appetite and abnormal clinical signs, such as pain, lameness, or swelling of the paws, were determined from the medical reports or with follow-up calls. Only 1 cat had mild swelling of the paws after the procedure, and it recovered the next day. No further side effects were observed during the procedure.

Three cats had been speculated to have concurrent diseases. Cat 1 was diagnosed with right cardiomegaly and pulmonary hypertension, and chylothorax gradually resolved within 2 months after administration of pimobendan (Vetmedin) and sildenafil (Viagra). Cat 6 had multiple large abdominal masses with concurrent chylothorax and peritoneal effusion. Granulomatous disease as in feline infectious peritonitis or lymphoma was suspected by means of cytologic evaluation of the masses, and coronavirus was detected in the pleural fluid; the cat’s clinical condition deteriorated, and it died 2 weeks after the CT examination. The TD enhancement failed in cat 7. In this cat, a mass at the level of the right third mammary gland and extremely enlarged ipsilateral inguinal and bilateral medial iliac lymph nodes were noted on the CT images; thus,
There was no clear effusion after surgery and tapped from 33 to 1 mL/d in 1 to 2 weeks. Thoracic tubes were placed in all 3 cats for monitoring postoperative pleural effusion, and all cats had no effusion postoperatively, and all cats had no effusion postoperatively. There were no complications or recurrence in the cases treated with surgery after 3 to 9 months of follow-up. Cat 5 with idiopathic chylothorax was lost to follow-up after the CT examination. The details of each cat are listed elsewhere (Supplementary Appendix S2).

Discussion

The present study demonstrated CT lymphangiography via intrametaatarsal pad injection in cats with chylothorax as a simple procedure with a high success rate (6 of 7 cats) within 5 to 15 minutes of injection. This approach allows for identification of lymphatic vessels, cisterna chyli location, and the variable TD branch locations. Moreover, successful migration of the contrast medium from the injection site to the lymphatic vessels occurred in all cases within 5 minutes. The recommended procedure is similar to that in a previous report in dogs, as follows: prepare at least 1 mL of contrast medium/kg (0.5 mL/kg/pad) and inject into the thickest region of the metatarsal pads at the same time; thoroughly and vigorously massage the pads and hind limbs for 3 to 4 minutes, then start the first CT scan within 5 minutes after initiating the injection. Failure to opacify the TD by the first scan means that serial delayed scans at 2- to 3-minute intervals may be necessary until a satisfactory TD enhancement is achieved.

The present study demonstrated a high success rate (86%) of CT lymphangiography via intrametaatarsal injection in cats, comparable with the success rate in dogs (90%). Previous reports have demonstrated differing success rates of ultrasound-guided intranodal (popliteal or mesenteric lymph nodes) injection for CT lymphangiography in cats (71% to 100%). Difficulty in the injection of the contrast medium into the popliteal lymph nodes and extranodal contrast leakage in the abdominal cavity via intramesenteric lymph node injection have also been described. In comparison, SC injection of contrast medium is less technically demanding and shows no abdominal leakage, either from the perianal region, dorsal metatarsal region, or intrametaatarsal pads. Moreover, successful migration of the contrast medium from the injection site to the lymphatic vessels occurred in all cases within 5 minutes. The recommended procedure is similar to that in a previous report in dogs, as follows: prepare at least 1 mL of contrast medium/kg (0.5 mL/kg/pad) and inject into the thickest region of the metatarsal pads at the same time; thoroughly and vigorously massage the pads and hind limbs for 3 to 4 minutes, then start the first CT scan within 5 minutes after initiating the injection. Failure to opacify the TD by the first scan means that serial delayed scans at 2- to 3-minute intervals may be necessary until a satisfactory TD enhancement is achieved.

The present study demonstrated a high success rate (86%) of CT lymphangiography via intrametaatarsal injection in cats, comparable with the success rate in dogs (90%). Previous reports have demonstrated differing success rates of ultrasound-guided intranodal (popliteal or mesenteric lymph nodes) injection for CT lymphangiography in cats (71% to 100%). Difficulty in the injection of the contrast medium into the popliteal lymph nodes and extranodal contrast leakage in the abdominal cavity via intramesenteric lymph node injection have also been described. In comparison, SC injection of contrast medium is less technically demanding and shows no abdominal leakage, either from the perianal region, dorsal metatarsal region, or intrametaatarsal pads. Moreover, successful migration of the contrast medium from the injection site to the lymphatic vessels occurred in all cases within 5 minutes. The recommended procedure is similar to that in a previous report in dogs, as follows: prepare at least 1 mL of contrast medium/kg (0.5 mL/kg/pad) and inject into the thickest region of the metatarsal pads at the same time; thoroughly and vigorously massage the pads and hind limbs for 3 to 4 minutes, then start the first CT scan within 5 minutes after initiating the injection. Failure to opacify the TD by the first scan means that serial delayed scans at 2- to 3-minute intervals may be necessary until a satisfactory TD enhancement is achieved.

The present study demonstrated a high success rate (86%) of CT lymphangiography via intrametaatarsal injection in cats, comparable with the success rate in dogs (90%). Previous reports have demonstrated differing success rates of ultrasound-guided intranodal (popliteal or mesenteric lymph nodes) injection for CT lymphangiography in cats (71% to 100%). Difficulty in the injection of the contrast medium into the popliteal lymph nodes and extranodal contrast leakage in the abdominal cavity via intramesenteric lymph node injection have also been described. In comparison, SC injection of contrast medium is less technically demanding and shows no abdominal leakage, either from the perianal region, dorsal metatarsal region, or intrametaatarsal pads. Moreover, successful migration of the contrast medium from the injection site to the lymphatic vessels occurred in all cases within 5 minutes. The recommended procedure is similar to that in a previous report in dogs, as follows: prepare at least 1 mL of contrast medium/kg (0.5 mL/kg/pad) and inject into the thickest region of the metatarsal pads at the same time; thoroughly and vigorously massage the pads and hind limbs for 3 to 4 minutes, then start the first CT scan within 5 minutes after initiating the injection. Failure to opacify the TD by the first scan means that serial delayed scans at 2- to 3-minute intervals may be necessary until a satisfactory TD enhancement is achieved.

The present study demonstrated a high success rate (86%) of CT lymphangiography via intrametaatarsal injection in cats, comparable with the success rate in dogs (90%). Previous reports have demonstrated differing success rates of ultrasound-guided intranodal (popliteal or mesenteric lymph nodes) injection for CT lymphangiography in cats (71% to 100%). Difficulty in the injection of the contrast medium into the popliteal lymph nodes and extranodal contrast leakage in the abdominal cavity via intramesenteric lymph node injection have also been described. In comparison, SC injection of contrast medium is less technically demanding and shows no abdominal leakage, either from the perianal region, dorsal metatarsal region, or intrametaatarsal pads. Moreover, successful migration of the contrast medium from the injection site to the lymphatic vessels occurred in all cases within 5 minutes. The recommended procedure is similar to that in a previous report in dogs, as follows: prepare at least 1 mL of contrast medium/kg (0.5 mL/kg/pad) and inject into the thickest region of the metatarsal pads at the same time; thoroughly and vigorously massage the pads and hind limbs for 3 to 4 minutes, then start the first CT scan within 5 minutes after initiating the injection. Failure to opacify the TD by the first scan means that serial delayed scans at 2- to 3-minute intervals may be necessary until a satisfactory TD enhancement is achieved.
most frequently used dose of contrast medium in our study was 1 mL/kg, similar to the dose used for intrametatarsal injection in dogs. \textsuperscript{20} Among the 6 cats with successful TD enhancement in the present study, grade 3 enhancement (4/6 cases) was achieved with contrast medium doses of 1 mL/kg (n = 2), 1.5 mL/kg (1), and 2 mL/kg (1), whereas grade 2 enhancement (2/6 cases) was achieved with a contrast medium dose of 1 mL/kg (1) and 2 mL/kg (1). One cat received an injection of 1.5 mL of contrast medium/kg, but no TD enhancement was achieved. A higher dose of contrast medium did not guarantee better quality of TD visualization, similar to the findings of a previous report\textsuperscript{20} in dogs. Thus, an initial dose of 1 mL of contrast medium/kg is recommended for intrametatarsal pad injection in cats.

The optimal timing for TD enhancement ranged from 5 to 15 minutes in our study, similar to intrametatarsal pad injection in dogs and intrapopliteal lymph node injection in cats.\textsuperscript{20,22} In our experience, thoroughly massaging the pads and hind limbs increases the success rate and reduces the time required for TD enhancement. Massaging the injection site is also recommended for perianal subcutaneous injection of contrast medium.\textsuperscript{16,17} However, the relatively concentrated accumulation of contrast medium in the pads may result in pad and limb massage being more efficient and practical.

The combination of TD ligation, subtotal pericardiectomy, and cisterna chyli ablation or omentalization followed by placement of pleural ports has been the most common approach for treating cats with chylothorax.\textsuperscript{14,15} Therefore, preoperative lymphangiography is recommended for a complete understanding of the morphology of the TDs or cisterna chyli.\textsuperscript{30} Previous literature suggests a left-sided approach for TD ligation in cats and a right-sided approach in dogs with chylothorax.\textsuperscript{11,10,31} However, in our study, 72% of the TD branches were located in the right hemithorax, similar to that (78% to 87%) seen in dogs but inconsistent with previous reports of cats.\textsuperscript{17,20,21,12,32} Furthermore, most of the TDs at the T11 to T12 level had only 1 or 2 branches, making complete ligation at this site easier. Thus, a right-sided approach for TD ligation may also be considered in cats. In fact, the 3 cats that received surgery exhibited right or dorsal TD branches at the T10 to T12 level in the preoperative CT lymphangiography, and complete TD ligation was achieved by means of a right-sided approach. Our results revealed the potential variation of TD morphology in cats and highlighted the importance of preoperative CT lymphangiography to guide the selection of approach side for TD ligation.

In our study, the cisterna chyli was located at the L2 to L3 level, medial to the middle of the right kidney and dorsal to the proximal descending colon. Compared with canine cisterna chyli,\textsuperscript{20} which spans from the L2 to L5 level with the most common site being L3, the location of feline cisterna chyli has relatively less variation among individuals. In contrast, the structure of cisterna chyli varies individually, since 5 types of cisterna chyli were identified in the small number of cases in our study. A left-sided 9th or 10th intercostal approach for cisterna chyli ablation has been described in cats with idiopathic chylothorax.\textsuperscript{14} However, 5 of 6 cats in our study exhibited right-sided distribution of the cisterna chyli, indicating that a right-sided approach may be considered if preoperative CT lymphangiography is available. These results may be valuable to aid the selection of the best approach sites and corridor for cisterna chyli ablation, as suggested in a previous study of dogs.\textsuperscript{30}

The TD failed to opacify in only 1 cat in the present study. A mass at the right third mammary gland with severe enlargement and heterogeneous enhancement of the bilateral internal iliac, medial iliac, and inguinal lymph nodes and blurring margin of the retroperitoneal fat were noted in this cat. These image findings suggested a mammary gland tumor with lymphatic metastasis or lymphoid tumors. However, further diagnostic workup or biopsy were declined by the owner and a definitive diagnosis was not available. Accumulation of the contrast medium was noted in the enlarged lymph nodes, and we presumed that the abnormal lymph nodes affected the lymphatic flow. However, successful migration of the contrast medium from the metatarsal region to the lymphatic vessels was still achieved.

Three systems of the superficial lymph vessels (superficial lateral, superficial medial, and deep medial systems) in the pelvic limbs have been described in dogs, and the identification of dorsal lymph vessel pathway, which courses along the gluteal region to the abdominal cavity, could be enhanced via intrametatarsal pad injection.\textsuperscript{20,33} Despite the fact that cats have similar lymphatic pathways, their anatomic structure in CT lymphangiography has not been reported.\textsuperscript{34} The cats in the present study exhibited a higher frequency (66%) of the dorsal lymph vessel pathway than dogs (34%), possibly indicating a more complicated communication of superficial lymph vessels in cats.\textsuperscript{20} Inconsistent drainage pathway in different pelvic limbs was also noted in 3 cats in the present study, and a previous study demonstrated that this inconsistency might result in failure of CT lymphangiography via unilateral popliteal lymph node injection.\textsuperscript{22} Hence, bilateral intrametatarsal pad injection of contrast medium may cover most pathways of the hind limbs to increase the success rate of lymphatic vessel enhancement. Intrametatarsal pad injection may also be considered a possible method of staging the malignancy of pelvic limb disease.

The main limitations of this study were its retrospective design and small sample size. Multiple institutions were included in this study to increase the sample size; however, this resulted in different operators performing the procedures, which may have affected the results. Additionally, surgery was performed to confirm the number of TD branches in only 3 of 7 cases.

In conclusion, feline CT lymphangiography by intrametatarsal pad injection is a safe and effective method to enhance the TDs and cisterna chyli in cases of chylothorax. Future studies including a larger sam-
ple size are warranted to establish the morphology of the feline TD and verify possible complications.

Acknowledgments
The authors received no financial support for the research, authorship, or publication of this article. The authors declare no conflicts of interest with respect to the research, authorship, or publication of this article. We thank Dr. Yi-Hsuan Lee for helping with data acquisition.

References

Supplementary Materials
Supplementary materials are posted online at the journal website: avmajournals.avma.org