Synovium secretome as a disease-modifying treatment for equine osteoarthritis

Santiago Mejia Hernandez, DVM1; Laila Begum, PhD1; Laura E. Keller, PhD1; Qin Fu, PhD2; Sheng Zhang, MD2; Lisa A. Fortier, DVM, PhD1*

1Department of Clinical Sciences, Cornell University, Ithaca, NY
2Proteomics and Metabolomics Facility, Cornell University, Ithaca, NY
*Corresponding author: Dr. Fortier (laf4@cornell.edu)
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OBJECTIVE
To identify chondroprotective factors as potential disease-modifying osteoarthritis treatments using an unbiased, bottom-up proteomics approach.

SAMPLES
Paired equine cartilage explants and synovial membrane were collected postmortem from 4 horses with no history of lameness and grossly normal joints at necropsy.

PROCEDURES
Six groups were established: cartilage, synoviocytes, and cartilage + synoviocytes (coculture), all with or without interleukin (IL)-1β. The catabolic effect of IL-1β was verified by glycosaminoglycan (GAG) released from cartilage into media by 1,9-dimethyl-methylene blue assay and cartilage toluidine blue histochemistry. Conditioned media from cocultures with or with IL-1β were submitted for bottom-up proteomic analysis. Synoviocyte gene expression was evaluated using reverse transcription–quantitative PCR (RT-qPCR) for proteins of interest identified in the proteomics scan.

RESULTS
GAG content was retained in cartilage when in cocultures treated with IL-1β. Fourteen proteins of interest were selected from the proteomic analysis. From these 14 proteins, metalloproteinase inhibitor 3 precursor (TIMP3), tumor necrosis factor receptor superfamily member 11B (TNFRSF11B), insulin-like growth factor–binding protein 2 (IGFBP2), and alpha-2 macroglobulin (A2M) were selected for synoviocyte gene expression analysis by RT-qPCR. Gene expression of TIMP3 (P = .02) and TNFRSF11B (P = .04) were significantly increased in synoviocytes from cocultures treated with IL-1β compared to controls. Contrary to expectations based on protein expression, IGFBP2 gene expression (P = .04) was significantly decreased in IL-1β-stimulated coculture synoviocytes compared to control coculture synoviocytes. A2M gene expression in synoviocytes was not different between coculture groups.

CLINICAL RELEVANCE
The secretome from synoviocytes could provide a milieu of bioactive factors to restore joint homeostasis in osteoarthritis.

Osteoarthritis (OA) is a progressive and debilitating disease affecting most mammalian species. In humans, it is estimated that the socioeconomic impact of musculoskeletal conditions, including OA, account for 1.0% to 2.5% of the gross domestic product, or an estimated $233.5 billion in the United States, Canada, the United Kingdom, France, and Australia.1–3 In 2013, the total national arthritis-attributable medical care expenditures and earnings losses among adults with arthritis in the United States alone were $303.5 billion.3 In horses, joint disease is the most common case of lameness, resulting in morbidity for the equine patients and loss of income for owners and trainers.4–9 OA is commonly treated with medications such as oral NSAIDs, corticosteroids, and sodium hyaluronate; and biologics such as platelet-rich plasma, mesenchymal stromal cells, interleukin (IL)-1 receptor antagonist protein, and others.10–17 Most of these treatments decrease pain, but do not necessarily address the disease course of OA. To affect the progression of OA, cartilage catabolism needs to be inhibited.18,19

Previous in vitro experiments revealed that synovial tissue or synoviocytes mitigate the catabolic effects of IL-1β on cartilage when the cell types are cultured together.11,12,20–23 In these studies, synovium or synoviocytes decreased IL-1β-induced glycosaminoglycan (GAG) loss, cartilage degradation, and synovial membrane inflammation. These studies suggest that the secretome of synoviocytes...
contains 1 or more bioactive mediators that protect cartilage from the degradative effects of IL-1β. However, these synovium secretome factors have not been identified. In this study, we aimed to identify synovium secretome factors using an unbiased, bottom-up proteomics approach. Identifying 1 or more unique molecules, or the entire secretome, could then be explored further as disease-modifying OA drugs for human and veterinary patients.

Materials and Methods

Study design

Tissue samples were collected from horses that were euthanized using IV pentobarbital (~1 mL/4.5 kg) as part of other projects at the institution. Samples were collected postmortem, with Institutional Animal Care and Use Committee approval. No animal was receiving medication for at least 2 months prior to euthanasia. Broadsly, cocultures of cartilage and synoviocytes were treated with and without IL-1β. Conditioned media containing the synoviocyte secretome were subjected to proteomic analysis. Relative protein abundance was calculated for IL-1β-treated cocultures versus control cocultures. Gene expression for proteins of interest was verified in synoviocytes by reverse transcription–quantitative PCR (RT-qPCR).

Cartilage and synoviocyte cocultures

Cartilage and synovial membrane were harvested from the patellofemoral joints of 4 horses euthanized for reasons unrelated to joint disease or this study (age range, 7 months to 7 years). All joints were visually normal. Paired full-thickness cartilage explants and synoviocytes were isolated to establish cocultures with synoviocytes on the bottom of the well, and cartilage explants were suspended in the transwell as described previously. Cultures were established in Dulbecco’s Modified Eagle Medium with 10% fetal bovine serum, 25 mM 4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid, 50 μg/mL ascorbic acid, 50 μg/mL α-ketoglutaric acid, 300 μg/mL L-glutamine, 100 U/mL penicillin sodium, and 100 μg/mL streptomycin sulfate at 37°C with 5% carbon dioxide and 90% humidity. After 24 hours, media were replaced with serum-free medium ± recombinant equine IL-1β (10 ng/mL, R&D Systems). Media were again exchanged after 48 and 72 hours, and the resultant conditioned medium was collected. Protease inhibitors were added (Protease Inhibitor Cocktail tablets; Roche Holding AG), and samples were centrifuged at 350 X g for 3 minutes to remove particulate matter and stored at −80°C until proteomic and GAG analyses were performed. Cartilage explants were fixed in 10% neutral buffered formalin for toluidine blue histochemistry; synoviocytes were lysed for gene expression analyses.

Toluidine blue histochemistry

Three cartilage explants per culture well were fixed in 10% neutral buffered formalin for 24 hours, processed, embedded in paraffin, sectioned (5 μm), and stained with toluidine blue to evaluate glycosaminoglycan distribution.

Bottom-up proteomics

The aim of our study was to identify proteins secreted by synoviocytes that protect cartilage from the catabolic effects of IL-1β. Therefore, the proteomic analysis was designed a priori to identify proteins that were increased in IL-1β-treated cocultures relative to control cocultures. Absolute protein quantification was not performed. Proteins were identified using Orbitrap Fusion by FT-Q-IT mode for solution-based protein identifications and associated label-free quantitation (LFQ).

In-solution digestion was performed on an S-Trap microspin column (ProtiFi) according to protocol, as described previously, with slight modification. Twenty micrograms of the protein samples in 25 μL buffer consisting of 50 mM tetraethylammonium bromide (TEAB, pH 8.5), 6 M urea, 2 M thiourea, and 1% SDS were reduced with 10 mM Tris(2-carboxyethyl) phosphine hydrochloride for 1 hour at 34°C, then was alkylated with 50 mM iodoacetamide for 1 hour in the dark and quenched with a final concentration of 36 mM dithiothreitol. After quenching, 12% phosphoric acid was added to a final concentration of 1.2%, followed by a 1:7 dilution (volume/volume) with binding buffer (90% methanol, 0.1 M TEAB, pH 8.5). The samples were then loaded into the S-Trap microcartridges and centrifuged at 4,000 X g for 30 seconds, and washed three times with 150 μL binding buffer by centrifuge at 4,000 X g for 30 seconds. Digestion was performed with 2 μg trypsin (1:10 weight/weight) in 125 μL 50 mM TEAB (pH 8.5) added to the top of the spin column and incubated at 37°C overnight. After incubation, the digested peptides were eluted off the S-Trap column sequentially, with 80 μL each of 50 mM TEAB (pH 8.5) followed by 0.2% formic acid (FA) and 50% acetonitrile in 0.2% FA. Three eluted peptides washes were pooled together and evaporated to dryness using a Speedvac SC110 (Thermo Savant).

Protein identification was performed by nanoscale liquid chromatography coupled to tandem mass spectrometry (nanoLC-MS/MS) analysis. The tryptic digests were reconstituted in 85 μL of 0.5% FA for nanoLC-MS/MS analysis, which was carried out using an Orbitrap Fusion Tribrid mass spectrometer (Thermo-Fisher Scientific) equipped with a nanospray Flex Ion Source and coupled with a Dionex UltiMate 3000 RSLCnano system (Thermo-Fisher Scientific) equipped with a nanoC18 RP trapping column (5 μm, 100 μm internal diameter X 20 mm) at a 20-μL/minute flow rate for rapid sample loading, and then separated on a PepMap™ C-18 reversed-phase (C-18 RP) nanocolumn (2 μm, 75 μm X 25 cm at 35°C). The peptides were eluted in a 90-minute gradient of 7% to 38% acetonitrile (ACN) in 0.1% FA at 300 nL/minute, followed by an 8-minute ramping to 90% ACN−0.1% FA and an 8-minute hold at 90% ACN−0.1% FA. The column was re-equilibrated with 0.1% FA for 25 minutes prior to the next injection. The Orbitrap Fusion
was operated in positive ion mode with the spray voltage set at 1.6 kV and the source temperature set at 275 °C. External calibration for FT, IT, and quadrupole mass analyzers was performed. For the data-dependent acquisition analysis, the instrument was operated using an FT mass analyzer and MS scan to select precursor ions, followed by 3-second “Top Speed” data-dependent collision-induced dissociation ion trap MS/MS scans at 1.6 m/z quadrupole isolation for precursor peptides with multiple charged ions above a threshold ion count of 10,000 and a normalized collision energy of 30%. MS survey scans were set at a resolving power of 120,000 50% of the maximum peak height (full width at half maximum) at m/z 200, for the mass range of m/z 375 to 1,575. Dynamic exclusion parameters were set at 50 seconds of exclusion duration with ± 10 ppm exclusion mass width. All data were acquired using Xcalibur 3.0 operation software (Thermo Fisher Scientific).

Data analysis

The data-dependent acquisition raw files for collision-induced dissociation MS/MS were subjected to database searches using Proteome Discoverer (PD) 2.3 software (Thermo Fisher Scientific) with the Sequest HT algorithm. The PD 2.3 processing workflow containing an additional node of the Minora Feature Detector for precursor ion-based quantification was used for protein identification and protein relative quantitation analysis between samples. The database search was conducted against an Equus caballus database containing 245,782 sequences downloaded from the National Center for Biotechnology Information. Two missed trypsin cleavage sites were allowed. The peptide precursor tolerance was set to 10 ppm; fragment ion tolerance was set to 0.6 Da. Variable modification of methionine oxidation, deamidation of asparagine/glutamine, N-terminal acetylation, and fixed modification of cysteine carbamidomethylation were set for the database search. Only high-confidence peptides defined by Sequest HT with a 1% of false discovery rate by Percolator were considered for the peptide identification. The final protein identifications contained protein groups that were filtered with at least 2 peptides per protein. Relative quantitation of identified proteins between the IL-1β-treated and control conditioned media samples was determined by the LFQ workflow in PD 2.3. The precursor abundance intensity for each peptide identified by MS/MS in each sample was determined automatically, and the unique plus razor peptides for each protein in each sample were summed and used for calculating the protein abundance by PD 2.3 software. Protein ratios were calculated based on a pairwise ratio for treated versus control samples.

Search Tool for the Retrieval of Interacting Genes/Proteins

Uniprot accession numbers of proteins of interest were subjected to database searches using the Search Tool for the Retrieval of Interacting Genes/Proteins (STRING) for visualization of the network between proteins, with an abundance ratio > 1 in coculture media with or without IL-1β. STRING (version 11.5, August 12, 2021; ELIXIR’s Core Data Resources) uses biologic databases from numerous sources, and Web resources of known and predicted protein–protein interactions.27

Synoviocyte gene expression

Synoviocytes were lysed, and total RNA was isolated using the RNeasy Mini Kit (QIAGEN). Purity and concentration of the RNA was assessed by UV microspectrophotometry (NanoDrop 1000 Spectrophotometer; Thermo Scientific). Based on proteomic results, gene expression of insulin-like growth factor-binding protein 2 (IGFBP2), metalloproteinase inhibitor 3 precursor (TIMP3), and tumor necrosis factor receptor superfamily member 11B (TNFRSF11B) were quantified by RT-qPCR using equine-specific primers and probes (Supplemental Table 1) and the Taqman One-Step RT-qPCR technique (ViiA 7 Real-Time PCR System, Applied Biosystems). The copy number of messenger RNA was determined using absolute qPCR derived from a standard curve developed for each gene at the time of analysis, and these values were normalized to the reference gene, 18S.

Statistical analysis

To compare gene expression in synoviocytes between IL-1β-treated and control cocultures, a paired sample t test was performed. A P value < .05 was considered significant.

Results

Toluidine blue histochemistry

To verify that synoviocytes protected cartilage from the catabolic effects of IL-1β, matrix metachromasia as an indicator of GAG content was examined in cartilage explants. In cartilage-only control cultures, matrix GAG was intense and evenly distributed throughout the explant (Figure 1). As anticipated, addition of IL-1β resulted in marked GAG loss, with complete GAG depletion in the superficial two thirds of the explants and with minimal GAG remaining in the deeper one third of the explants. In cartilage cocultured with synoviocytes, matrix metachromasia was similar or increased compared to cartilage-only cultures, and when cocultures were treated with IL-1β, GAG staining was similar to cartilage-only controls in the deeper layers, with GAG loss confined to the superficial one third of the explant. These results confirmed that the presence of synoviocytes protected cartilage from IL-1β-induced matrix GAG loss. Spectrophotometric quantification of GAG loss was not performed because the goal of our study was to perform proteomic analysis. Histologic confirmation of IL-1β-induced cartilage matrix GAG loss confirmed that the culture conditions were appropriate to continue with proteomic analysis.

Proteomic analysis of conditioned media

The aim of our study was to identify proteins secreted from synoviocytes that protected cartilage...
from IL-1β-induced catabolism, so only conditioned media from cocultures were used for proteomics. In conditioned media, 639 proteins met the criteria of at least 2 unique peptides with 95% confidence. To identify proteins that were upregulated in IL-1β-stimulated cocultures, an abundance ratio was calculated as protein “X” in cocultures with IL-1β versus protein “X” in cocultures without IL-1β. Unique proteins (n = 132) with an abundance ratio > 1 were identified (Supplemental Table 2) and classified by their molecular function as extracellular matrix, 51%; structural, 18%; catabolic, 15%; extracellular matrix/structural, 12%; chemokine, 2%; or growth factor, 2%. Structural proteins such as keratin, collagen, and actin were deemed of less interest, iterating the list to 14 proteins of interest (Table 1). Functional protein–protein association networks were assessed using STRING analysis (Figure 2). From these 14 proteins, based on a subjective combination of abundance ratios, known role in cartilage biology, and the STRING analysis, TIMP3, IGFBP2, TNFRSF11B (aka osteoprotegerin [OPG]), and alpha-2 macroglobulin (A2M) genes were selected to be measured in synoviocytes by qPCR.

**Synoviocyte gene expression**

TIMP3 is a potent inhibitor of matrix-degrading aggrecanases and collagenases. Synoviocyte TIMP3 expression (P = .02) in cocultures treated with IL-1β increased ~3-fold compared to cocultures without IL-1β (Figure 3).

TNFRSF11B encodes for the decoy receptor OPG and functions to suppress chondrocyte apoptosis by binding to and suppressing apoptosis in chondrocytes induced by tumor necrosis factor-related apoptosis-inducing ligand (TRAIL). Synoviocyte TNFRSF11B expression (P = .04) in IL-1β-stimulated cocultures increased ~10-fold compared to control cocultures (Figure 3). IGFBPs are involved in the transport of IGF into cartilage, and are associated with increased production of proteoglycans by chondrocytes and can inhibit chondrocyte apoptosis. In contrast to the protein abundance ratio, synoviocyte IGFBP2 expression was decreased by

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**Table 1**—Proteins of interest with putative chondroprotective function were identified based on the abundance ratio of interleukin (IL)-1β-treated to control cocultures and molecular function.

<table>
<thead>
<tr>
<th>Protein</th>
<th>Abbreviation</th>
<th>Average abundance ratio</th>
<th>NCBI accession no.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Metalloproteinase inhibitor 3 precursor</td>
<td>TIMP3</td>
<td>4.493</td>
<td>126352371</td>
</tr>
<tr>
<td>Insulin-like growth factor II</td>
<td>IGF2</td>
<td>4.171</td>
<td>133565611</td>
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<tr>
<td>Thrombospondin-1 precursor</td>
<td>THBS1 Precursor</td>
<td>2.247</td>
<td>822092742</td>
</tr>
<tr>
<td>Tumor necrosis factor receptor superfamily member 11B</td>
<td>TNFRSF11B</td>
<td>1.745</td>
<td>953862440</td>
</tr>
<tr>
<td>Vascular cell adhesion molecule</td>
<td>VCAM</td>
<td>1.697</td>
<td>154756883</td>
</tr>
<tr>
<td>Complement factor B</td>
<td>CFB</td>
<td>1.455</td>
<td>149732066</td>
</tr>
<tr>
<td>Afamin</td>
<td>AFM</td>
<td>1.449</td>
<td>149701611</td>
</tr>
<tr>
<td>C-C motif chemokine 16</td>
<td>CCL16</td>
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<td>133562957</td>
</tr>
<tr>
<td>Gelsolin</td>
<td>GSN</td>
<td>1.440</td>
<td>260656201</td>
</tr>
<tr>
<td>Insulin-like growth factor-binding protein 2</td>
<td>IGFBP2</td>
<td>1.419</td>
<td>133674713</td>
</tr>
<tr>
<td>Component C7</td>
<td>C7</td>
<td>1.394</td>
<td>133603815</td>
</tr>
<tr>
<td>Inter-alpha-trypsin inhibitor</td>
<td>ITIH3</td>
<td>1.363</td>
<td>194227183</td>
</tr>
<tr>
<td>Lumican</td>
<td>LUM</td>
<td>1.336</td>
<td>124377698</td>
</tr>
<tr>
<td>Alpha-2-macroglobulin</td>
<td>A2M</td>
<td>1.249</td>
<td>95385941</td>
</tr>
</tbody>
</table>

Abundance ratios were calculated based on pairwise ratio for a given protein in IL-β-treated coculture to control coculture samples. NCBI = National Center for Biotechnology Information. Proteins are listed in decreasing order of abundance ratio.
~0.2-fold ($P = .04$) in the presence of IL-1β compared to control cocultures, suggesting that IGFBP2 could be expressed by chondrocytes. Previous studies have shown that A2M can reduce catabolic damage in vitro and in preclinical post-traumatic OA models through its broad-spectrum ability to block catabolic cytokines and enzymes including IL-1β, thus downregulating the inflammatory cascade. There was no difference in A2M gene expression between synoviocytes in control cocultures ($1.1 ± 1.39$) and IL-1β-treated cocultures ($0.21 ± 0.08; P = .29$).
Discussion

This study was based on previous observations that synoviocytes protect cartilage from the catabolic effects of IL-1β in vitro.\textsuperscript{20,25,39,40} IL-1β is commonly used in vitro to stimulate cartilage catabolism and inflammation by inducing expression of matrix metalloproteinases (MMPs), a disintegrin and metalloproteinase with thrombospondin motifs (ADAMTS), and other catabolic enzymes.\textsuperscript{41} Similar to the aforementioned studies, cartilage treated with IL-1β resulted in GAG depletion; however, when in coculture with synoviocytes, GAG was mostly retained, confirming that the synovium secretome has chondroprotective properties against the catabolic effects of IL-1β.

TIMP3 was selected for gene expression analysis because it is an antagonist to many cartilage degradative enzymes such as MMPs, ADAMTS, and aggrecanases, and it has been shown to decrease ADAMTS-4-, ADAMTS-5-, and IL-1α-induced cartilage degradation.\textsuperscript{28} The abundance ratio of TIMP3 combined with the gene expression of TIMP3 in synoviocytes from IL-1β treated compared to control cocultures suggests that synoviocytes are synthesizing TIMP3, which could be leveraged for secretome drug development.

The protein TNFRSF11B is also known as OPG. The abundance ratio of TNFRSF11B and gene expression of TNFRSF11B was increased in synoviocytes in IL-1β-stimulated cocultures. This suggests that, like TIMP3, synoviocytes synthesize and secrete OPG. The role of OPG in OA is not completely understood; however, it is known to play important roles in bone turnover, cartilage homeostasis, and the onset of OA.\textsuperscript{62} OPG acts as a decoy receptor for the receptor activator of nuclear factor kappa-B ligand, preventing it from binding to the receptor activator of nuclear factor kappa-B on osteoblast precursors and driving their differentiation into osteoclasts. This suggests that aberrant OPG function can affect the balance between subchondral bone formation and resorption, making OPG essential in joint homeostasis.\textsuperscript{43–48} OPG has also been shown to slow the progression of OA effectively by suppressing chondrocyte apoptosis\textsuperscript{12,29} via binding to TRAIL.\textsuperscript{31,32,47} The chondroprotective effects of OPG are quite different and complementary to those of TIMP3, suggesting that optimizing the synoviocyte secretome would yield a more broadly therapeutic disease-modifying OA drug than targeting any single bioactive factor.

The abundance ratio of IGFBP2 was similar to the previously mentioned proteins; however, gene expression was decreased in synoviocytes in IL-1β-stimulated cocultures. This suggests that IGFBP2 in the coculture medium was not likely from the synoviocytes and may have diffused from the cartilage or was synthesized by chondrocytes. Insufficient cartilage was available for gene or protein expression analysis, and as this was not the aim of our study, it cannot be confirmed, but chondrocytes were the only other cell source in the culture system. Alternatively, the addition of IL-1β to the media could have affected synoviocyte IGFBP2 gene expression negatively, as suggested by a study\textsuperscript{49} in which IL-1β and IL-6 decreased IGFBP2 and IGFBP4 expression by epithelial cells in vitro.

The abundance ratio of A2M was 1.24, but there was no difference in expression between synoviocytes in coculture control and IL-1β-treated cocultures. Currently, A2M is showing promise as a negative regulator for many cartilage catabolic enzymes in both in vitro and in vivo models. When injected into an A2M knock-out mouse, A2M was primarily a serum protein and is produced by the liver. The results of our study suggest that A2M is synthesized by both chondrocytes and synoviocytes, but at a very low level, which is likely clinically insignificant.

In conclusion, our study provided proteomics-based evidence that the synovium secretome has the potential to protect articular cartilage from degradative effects of IL-1β by protein expression and by multiple interconnecting mechanisms, potentially through the decreased inflammatory pathways and the modulation of humoral and cellular immune responses. Our results suggest that there is more than 1 protein capable of protecting articular cartilage from the catabolic effects of IL-1β. The use of the proteomic method for determining the mechanisms behind the protective effects provided a comprehensive discovery of known and novel proteins secreted by synoviocytes with anabolic and catabolic properties involved in articular homeostasis. This study could be the starting point for future studies in which the synovium secretome factor could be evaluated in vivo and in vivo scenarios to assess the to validate the protective roles of the synovium secretome in OA as a disease-modifying product.

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The authors declare that there were no conflicts of interest. Dr. Fortier serves as Editor-in-Chief for the American Journal of Veterinary Research (AJVR). She declares that she had no role in the editorial direction of this manuscript.

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**Supplementary Materials**

Supplementary materials are posted online at the journal website: avmajournals.avma.org